

REMARKS

The Office Action has been carefully studied. No claim is allowed. Claims 1, 7-10, 12-16, 25, 28, 38, 39, 41, 42, 46, 48-50, 52, and 56-60 presently appear in this application and define patentable subject matter warranting their allowance. Reconsideration and allowance are hereby respectfully solicited.

The face-to-face interview among Dr. Wu, one of the present inventors, and the undersigned, representing applicants, and Examiners Dr. Kaushal and Dr. Fredman on August 3, 2004, is hereby gratefully acknowledged. Applicants' representatives wish to thank the examiners for the courtesies extended during this interview.

No specific prior art was discussed at the interview as there were no prior art issues raised in the Office Action. All the claims however were discussed, and applicants' representatives agreed to cancel the claims directed to a DNA molecule or convert them to method claims. Applicants' representatives further presented evidence on the interchangeability of uromodulin promoters among mammalian species and also proposed to limit the claims to "mammalian uromodulin promoters", to delete the recitation of "ancestors" from the claims to transgenic non-human mammals, and to incorporate the recitation of expression of the heterologous polypeptide "to the ascending limb of Henle's loop and to the early distal tubules of the kidneys" in method claim 38 in order to overcome the enablement rejection. Agreement was reached that the enablement rejection would likely fall.

The written description rejection was also discussed at the interview, with applicants' representatives arguing that there is a

sufficient representative number of uromodulin promoter sequences identified in the instant specification and in the prior art to satisfy the written description requirements. The only agreement reached on this written description issue was that applicants would introduce new claims directed to specific mammalian uromodulin promoters and specific transgenic non-human mammals and the examiners would carefully consider this issue in view of applicants' arguments in the instant amendment. The argument presented at the interview as it relates to the written description rejection is incorporated below.

Claims 41 and 55 have been rejected under 35 U.S.C. §112, first paragraph, as failing to comply with the written description requirement (new matter issues).

The recitation of "pig... uromodulin promoter" is now deleted from claim 41, thereby obviating this part of the rejection. Claim 41 is now amended to recite "human... uromodulin promoter" in place of pig uromodulin promoter as supported on page 19, lines 15-27 of the specification.

The feature of "mammalian uromodulin promoter directs expression of said heterologous polypeptide *in vivo* to the ascending limb of Henle's loop and the early distal tubules of the kidneys" as now recited in claims 38, 57 and 59 (claim 55 is now cancelled) is supported on page 14, lines 23-26 and page 16, lines 21-27.

Reconsideration and withdrawal of this rejection are therefore respectfully requested.

Claim 8 has been rejected under 35 U.S.C. §112, second paragraph, as being indefinite for the recitation of "predicted β -

turns". This rejection appears to no longer be an issue as the examiners have accepted applicants' argument that predicting the location of β -turns using common and publicly available algorithm, such as the Chou-Fasman algorithms, among others, is well recognized by those of skill in the art and therefore is not indefinite even if the specific algorithm is not mentioned by name in the specification.

Claims 1, 7-10, 12-16, 29, 31-37, 47 and 55 have been rejected under 35 U.S.C. §112, first paragraph, for lack of enablement. Claims 25, 28, 38-42, 44, 46, and 48-54 have also been separately rejected under 35 U.S.C. §112, first paragraph, for lack of enablement. Both of these rejections are obviated by the amendments to the claims as discussed at the face-to-face interview on August 3, 2004, the evidence on interchangeability of mammalian promoters as presented below in applicants' arguments rebutting the written description rejection, and the merely routine experimentation that is required by those of skill in the art to obtain transgenic non-human mammals.

Claims 1, 7-10, 12-16, 25, 28-29, 31-42, 44, and 46-55 have been rejected under 35 U.S.C. §112, first paragraph, as containing subject matter which was not described in the specification in such a way as to reasonably convey to one skilled in the relevant art that the inventor(s), at the time the application was filed, had possession of the claimed invention. This rejection is respectfully traversed.

The rejection as it relates to the scope of the transgenic animals and as discussed at the interview no longer appears to be at issue, particularly in view of the species of mammals recited in the claims being well described in the specification and further in view of

the examiner's agreement that enablement with regard to the different species of transgenic non-human mammals would be withdrawn as a result of the amendments to the claims.

The examiner holds that, with regard to uromodulin promoters, the instant specification only teaches the murine, goat and bovine uromodulin promoters but fails to disclose uromodulin promoter sequences and states, "Beside the mouse uromodulin promoter comprising the nucleotide sequences of SEQ ID NO:1, the specification as filed fails to disclose any other uromodulin promoter or fragment thereof (obtained from any amphibian, reptile, bird or all mammal) that directs the expression of a polypeptide *in vivo* in the kidney to produce the polypeptide in the urine."

In response to the examiner's position in this part of the rejection, applicants have now amended the claims to recite for only "mammalian" uromodulin promoters. As disclosed in the present specification at page 19, lines 15-18, the sequence of the mouse and goat uromodulin promoters have now been determined and reported by the applicants in the instant specification and the bovine, rat and human promoter regions have been previously reported in the prior art. On page 18, lines 15-19, a Yu et al. (1994) reference is cited for its disclosure of the bovine and rat (and human) uromodulin promoter regions and is furthermore incorporated by reference. This Yu et al. (1994) reference is the Yu et al., "Bovine and rodent Tamm-Horsfall protein (THP) genes-cloning, structural analysis, and promoter identification", Gene Expression 4:63-75 (1994) reference AM submitted with the Information Disclosure Statement dated July 13, 2000. For

clarification, uromodulin is also known as Tamm-Horsfall protein (THP). Fig. 5 in this Yu et al. (1994) reference shows a sequence alignment of the promoter regions of bovine, rat and human uromodulin. Accordingly, the uromodulin promoters from mouse, goat, cattle, rat, and human are disclosed in the instant specification and provides a representative number of species of the genus of mammalian uromodulin promoters so as to reasonably convey to those of skill in the relevant art that the inventors, at the time the application was filed, had possession of the claimed invention. This is further supported by the following discussion below.

Zbiskowska et al. (2002), cited by the examiner, discloses that a human uromodulin promoter provides kidney-specific expression in transgenic mice that leads to production of a recombinant biologically active human antitrypsin in the urine. Zbiskowska arbitrarily chose a human uromodulin promoter region that also contains exon 1 and part of exon 2 to drive expression, even though the exon sequences are unnecessary for kidney-specific expression. This can be seen from applicants' own evidence (Zhu et al., 2002 and 2003, copies of which were submitted with the amendment filed July 28, 2003), where sequences from only the 5'-upstream non-coding region were found to be sufficient for directing high level kidney-specific expression.

The Kim et al., Transgenic Res. 12:191-201 (2003) reference, which was also submitted with the amendment of July 28, 2003, demonstrates that the most critical cis elements of the uromodulin gene are located within the 600 bp upstream region (see abstract), providing evidentiary support for applicants' expectation in the specification at

page 18, lines 23-28, that the approximately 600 base pairs of sequence upstream of the uromodulin coding sequence is sufficient for directing kidney specific expression. Moreover, like Zbiskowska's showing of a human uromodulin promoter being capable of kidney specific expression in mice, Kim et al. also establish that a bovine uromodulin promoter can likewise function in mice. These results demonstrate the interchangeability of mammalian uromodulin promoters from one mammalian species to another, which is altogether expected given the disclosure in the specification at page 17, lines 13-15, that uromodulin promoter sequences are likely to be conserved among mammals and the sequence conservation actually observed among the disclosed bovine, rat, human, mouse and goat uromodulin promoter sequences.

To extend the alignment for bovine, rat and human uromodulin promoter sequences in Fig. 5 of Yu et al. (1994), applicants have prepared a comparison/alignment of about 600 base pairs upstream of the uromodulin coding sequence from all five bovine, rat, human, mouse and goat uromodulin promoters. The first alignment attached hereto for the examiner's consideration is entitled "Highly conserved nature of the proximal promoter sequences of rat, mouse, cattle, goat and human uromodulin (Tamm-Horsfall protein)" and was generated using a PileUp program of an online sequence analysis software (SeqWeb). A consensus sequence, shown as the bottom line in the alignment, is readily generated from the alignment of uromodulin promoter sequences from the five mammalian species. The second sequence alignment attached hereto shows the conserved transcription factor binding sites in the proximal promoters of uromodulin.

Also attached hereto are computer printouts of sequence identity calculations (BestFit) between the approximately 600 bp uromodulin promoter sequences presented in the first and second sequence alignments. A table summarizing the sequence identity calculated between each of five uromodulin promoter sequences is provided as a further attachment for the examiner's consideration. This table shows that the level of sequence identity is exceedingly high for promoters, ranging from 73% to 94% over approximately 600 base pairs.

On page 14, lines 17-20 of the instant specification, it is disclosed that uromodulin (Tamm-Horsfall protein) is by far the most abundant urinary protein of human and higher mammals, with an excretion rate of up to 200 mg per day.

In conclusion, when those of skill in the art take the five disclosed mammalian uromodulin promoters altogether into consideration along with (1) the high sequence identity between the five identified uromodulin promoters (sufficient to generate a consensus sequence), (2) the expectation that other mammalian promoters would have similar high sequence identity to the five identified mammalian uromodulin promoters and the consensus mammalian uromodulin promoter, (3) the functional interchangeability of the mammalian promoters among mammalian species, and (4) the ease in readily identifying a uromodulin promoter based not only on its sequence (i.e., sequence identity) but also on its ability to drive the expression of the most abundant (and therefore easily detectable) urinary protein in mammals for secretion into the urine at the ascending limb of Henle's loop and the early distal tubules of the

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kidneys, these same persons of skill in the art would immediately recognize that the inventors, at the time the application was filed, had possession of the claimed invention.

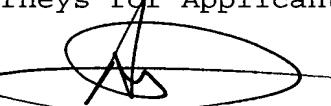
Reconsideration and withdrawal of the rejection are therefore respectfully requested.

In view of the above, the claims comply with 35 U.S.C. §112 and define patentable subject matter warranting their allowance. Favorable consideration and early allowance are earnestly urged.

Respectfully submitted,

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**HIGHLY CONSERVED NATURE OF THE PROXIMAL PROMOTER SEQUENCES
OF RAT, MOUSE, CATTLE, GOAT AND HUMAN UROMODULIN (TAMM-HORSFALL PROTEIN)**

RAT	1	31
MOUSE	C TAGTCTTGT.	CTGACAGAGG TCCAGTTGAG
CATTLE	C CAGTCTTGT.	CTGACAGAGG TCCAGTTCA
GOAT	C TGTTCCAATG	ATGTCTGAAT TGCCTCTGT
HUMAN	C TGTTCCAATG	ATGTCTGAAT TATCTGCTGT
CONSENSUS	C AGGTCCAGTG	ATGTCTGAAC TACCTCTGG
	C TGTTCCAAGTG	ATGTCTGAA- T-CCTCTGG
RAT	32	81
MOUSE	GGATGTCCAG ATGGTCTTGC	AACC.GATAA CTTTCTCAGA GACTCTCT
CATTLE	GAGTGTCCAG ATGGTCTGAT	AACCTGATGC CATTCTCAGA GA....CTCT
GOAT	CTCTGACCTT CAGGCATTCT	CAGCTCCTT CCTGCTCACA TCGGGACCCC
HUMAN	CTCTGACCTT CAGCCATTCT	CAGCTCCTT CCTGATCACA TTGGGACCCC
CONSENSUS	TTCTGACTTT CAGGCCATTCT	CAGCTCCTCT CTTGCTTGTG TCTGGATTCT
	-TCTGACCTT CAGGCATTCT	CAGCTCCTT C-TGCTCA-A T-GGGACTCT
RAT	82	131
MOUSE	TTCCTGTCTG GACTCTAGTG	GGGAGGACTA ATCTGGTCAA GCTGTTCTTC
CATTLE	TTCCTGTCTG GAATCTAGTG	AGGAGGACTT ATCTGGTCAA GCTGTCCTTT
GOAT	AGGGAAAGCTG GTTGAACCCA	TGAGGATGGA ACTTGCTTGT GAACTGAGTG
HUMAN	AGGGGAGCTG GCTGAATCTG	TGAGGATGGC ATTGCTTGT GAATTAAGTG
CONSENSUS	AAGGCTGATC TCATGAGAAT	GGGTGTTCA GAAGGGTGCC CTCTCCA...
	A-GGT-GCTG G--T-AA-TG	-GG-G-T--A AT-TGGTGG-- G---TCA-TG
RAT	132	181
MOUSE	AGATCAGGTG TGTGTTCCAG	GCTTCGAAGC AAATGTTCT GTTATCCTAA
CATTLE	AGAACAGGAG TGTGTTCCAG	TCTTCAAAGC AAACATTCTC TTTATCCTAA
GOAT	GCCACAAGTA TACATCCCAG	TGGGGACAGT GAGCACCCCT TTTCTCCTGG
HUMAN	GCCACAAGTA CACATCCTGG	TGGGGACGAT GAGCACCCCT TTTCTCCTGG
CONSENSUS	.AGACAGGTG CACCTCCCAT	CTGGGGCAGT GAATA.TCCT TTTGTCCTTA
	--ACAGGTG TAC-TCCCAG	T-GGGACAGT GAACA-TCCT TTT-TCCT-A
RAT	182	231
MOUSE	CCCAGGCTGG CTTCAGATAT	..TGTCTTTT TTCTGGCCCC TTTGG..TAT
CATTLE	CACAGTCTGA CTTCAGATAT	ACTGTCTTTT TCCTGGCTCC TTGGGCTTAG
GOAT	AGCAGCCTGG CTTCAGATT.	.CTGGCCTCT GCTT...T..
HUMAN	AGCAGCCTGG CTTCAGATT.	.CTGGCCTCT GCTTGGCT..
CONSENSUS	TGCAGCCTGG CTTCAGATA.	.CTGGCCTCT GCCTGGCTCC TTG.....A
	-GCAGCCTGG CTTCAGATAT	-CTGGCCTCT GCCTGGCTCC TTGGG--TA-
RAT	232	281
MOUSE	TTCCACCTTG TCCTTGCCCA	GGTCCAAGAA AAAGCCCAGA ACCTTGGCAC
CATTLE	GTCTACCTTG TCCTTGCCCA	GGTCCAAGAA AAGGCCAGA ACCTTGGCAC
GOAT	..CCACTTTG TGCTTTCAA	TGACCCAAGAA .AAGCCCAGG CACTTGGAAAT
HUMAN	..CCACTTTG TGCTTTCAA	TGACCCAAGAA .AATCCAGG CCCTTGGAAAT
CONSENSUS	TCCCACCTG CCCTTGTCA	TGACCCAAGAA GAAGGCCAGC ACCTTGGCAC
	TTCCACCTTG TCCTTGCAA	TGACCCAAGAA AAAGCCCAG- ACCTTGGCAC

	282	
RAT	TGCTTGCCTA GTTAATGTCT AACCGAGGAA TGTCTTGCTG CCAAAAGGTG	
MOUSE	TGTTTGCCCA GTTAATGTCT AACTGAGGAA TGTCTTGCTG CCAAAAGGTG	
CATTLE	TTTTTACCCA GTTAATTCTC AACTAAAGAA CCTCTCGTTG CCAAAAGATA	
GOAT	TGTTTACTCA GTTAATTCTC AACTAAAGAA CCTCTTGTTG CCAAAAGGTA	
HUMAN	TGCTTCCCA GTTAATTCTC AACTATGGAA TCTCTTGCTG TTAGAAGGTG	
CONSENSUS	TGTTTCCCA GTTAATTCTC AACTAAGGAA TCTCTTGCTG CCAAAAGGTG	
	332	381
RAT	.CAAACAGAG ACCTTGTATT TCCAGGCACA GGTGTGACCC C CAATGTCA	
MOUSE	.AAAACAGAG ACCTTGTATT TCCAGGCACA GGTGTGACCC C AATGTCA	
CATTLE	TAACACAGAG CCCTTGTAAAC TCTGGGCACA ACTGTGACCC C AGTGTCA	
GOAT	TAACACAGAG CCCTTGTAGC TGTGGGCACA GCTGTGACCC C CATGTCA	
HUMAN	CGAAACAGTG ACCTTGTATT TCCGGGCACA GGTGTGACCC CCCAATGTCA	
CONSENSUS	TAACACAGAG ACCTTGTATT TCCGGGCACA GGTGTGACCC C-CAATGTCA	
	382	431
RAT	ATCATTTCCT GTCTCTAACT ACCAGAGGAA AAACTAACAA CAACAGCCTC	
MOUSE	ATCATTT..T GTGTCTAACT CCCAGGGAA AAACTAACAA CAACAGACTC	
CATTLE	ATCATTGGG GTCTCTACCT ATTAG GGAA AA GAACAA CAACCACCTC	
GOAT	ATCATTGGG GTCTCTACCT ATTAG GGAA AA GAACAA CANCCACCTC	
HUMAN	ATCATTGGG GTCTCTAGCT ATTA GGAA AA AGAACAA CAACAACCTC	
CONSENSUS	ATCATTGGG GTCTCTA-CT ATTAG-GGAA AAAAGAACAA CAACAACCTC	
	432	481
RAT	ATGGTTTGGAA AAAGGTGAAC TCTATGCCAA ATGGGAAGAA AAATTCTGAC	
MOUSE	ATGGCTTGGAA AAAGGTGAAT TCTATGCCAA AAGGGAGGA AAGTTCT.AC	
CATTLE	ACAGCCTAGA AAAGGAAAAC ACTGTGCAA AAGGGAAAAA TATTCC..AC	
GOAT	ACAGCCTANA AAAGGAAAAC ACTGTGCAA AAGGGAAAAA TATTCC..AC	
HUMAN	ACAGCTTGGAA CAAGGCAAAC ATTATGCCAG GAGGAAAAAA TATTCC..AC	
CONSENSUS	ACAGCTTGGAA AAAGG-AAAC ACTATGCCAA AAGGGAAAAA TATTCC-AC	
	482	531
RAT	CCCCCACAGAA ACAATCTCAA GAGGCAGAAG CAGAGAATAA TTGGAGG.GA	
MOUSE	CCCCCACAGAA ACAATCTCAG AGGGCAGAAG CAGAGAATAA TCTGAGG.GA	
CATTLE	CCCCATTAAGATA..ATTAA GAAACAGAAC CAGAGGATCA TTGGAGGAAA	
GOAT	CCCCATTAAGATA..ATTAA GAAACAGAAC CAGAGGATCA TTGGAGGAGA	
HUMAN	CCCCAAGAAA ACAATATCAA AAAACAGAAC TAGAGACTAA TTGGAGGAGA	
CONSENSUS	CCCCA--AAA ACAATATCAA GAAACAGAAC CAGAGAATAA TTGGAGGAGA	
	532	581
RAT	GAGGGCCAGC CAAGGGCAGA CATATATATA TATATATTGA TCACAGGCAC	
MOUSE	GAGGGCCAGC CAAGGGCAGG CAAGTATATA TTGA TCACAGGCAC	
CATTLE	GAATGCCAGT GGGGGACAGA TGTATATATA TAGATATGAT AGTCACCTAC	
GOAT	GATTGCCAGT GGGGGACAGA TGTATATATA TAGATATGAA AGTCACCTAC	
HUMAN	GATTGCCAGC CTGGGGCAAA TGTGTATATA TAAGTATGAG GCACA.....	
CONSENSUS	GA-TGCCAGC C-GGGGCAGA TGTATATATA TA-ATATGAA -CACA---AC	
	582 591	
RAT	TTACTTGTGA	
MOUSE	TTACTTGTGA	
CATTLE	TTGTAAAAGG	
GOAT	TTGTAAAAGG	
HUMAN	TCATCACCAAG	
CONSENSUS	TTAT-A--GG	

**CONSERVED TRANSCRIPTION FACTOR BINDING SITES IN THE
PROMIXMAL PROMOTERS OF UROMODULIN**

RAT	1	31
MOUSE	C TAGTCTTGT. CTGACAGAGG TCCAGTTGAG	
CATTLE	C CAGTCTTGT. CTGACAGAGG TCCAGTTCAG	
GOAT	C TGGTCCAATG ATGTCTGAAT TGCCTCTGT	
HUMAN	C TGGTCCAATG ATGTCTGAAT TATCTGCTGT	
CONSENSUS	C AGGTCCAGTG ATGTCTGAAC TACCTCTGG	
	C TGGTCCAAGTG ATGTCTGAA- T-CCTCTGG	
RAT	32	81
MOUSE	GGATGTCCAG ATGGTCTTGC AACCA.GATAA CTTTCTCAGA GACTCTCT	
CATTLE	GAGTGTCCAG ATGGTCTGAT AACCTGATGC CATTCTCAGA GA...CTCT	
GOAT	CTCTGACCTT CAGGCCATTCT CAGCTCCTT CCTGCTCAC A TCAGGACCCC	
HUMAN	CTCTGACCTT CAGGCCATTCT CAGCTCCTT CCTGATCAC A TTGGGACCCC	
CONSENSUS	TTCTGACTT CAGGCCATTCT CAGCTCCTCT CTTGCTTGTG TCTGGATTCT	
	-TCTGACCTT CAGGCCATTCT CAGCTCCTT C-TGCTCA-A T-GGGACTCT	
RAT	82	131
MOUSE	TTCCTGTCTG GACTCTAGTG GGGAGGACTA ATCTGGTGAA GCTGTTCTTC	
CATTLE	TTCCTGTCTG GAATCTAGTG AGGAGGACTT ATCTGGTGAA GCTGTCCTT	
GOAT	AGGGAAGCTG GTTGAACCCA TGAGGATGGA ACTTGCTTG GAACTGAGTG	
HUMAN	AGGGGAGCTG GCTGAATCTG TGAGGATGGA ATTGCTTG GAAATTAAAGTG	
CONSENSUS	AAGGCTGATC TCATGAGAAT GGGTGTTCAGA GAAGGGTGCC CTCTCCA...-	
	A-GGT-GCTG G--T-AA-TG -GG-G-T--A AT-TGGTG-- G---TCA-TG	
RAT	132	TCF-1 181
MOUSE	AGATCAGGTG TGTGTTCCAG GCTTCGAAGC AAATGTTCT GTTATCCTAA	
CATTLE	AGAACAGGAG TGTGTTCCAG TCTTCAAAGC AAACATTCTCCT TTTATCCTAA	
GOAT	GCCACAAGTA TACATCCCAG TGAGGACAGT GAGCACCCCT TTTCTCCTGG	
HUMAN	GCCACAAGTA CACATCCTGG TGAGGACAGT GAGCACCCCT TTTCTCCTGG	
CONSENSUS	.AGACAGGTG CACCTCCCAT CTGGGGCAGT GAATA.TCCT TTTGTCCCTA	
	---ACAGGTG TAC-TCCCAG T-GGGACAGT GAACA-TCTCCTA TTT-TCCT-A	
RAT	182	NF-GMB 231
MOUSE	CCCAGGCTGG CTTCAGATAT ..TGTCTTTT TTCCTGCCCT TTTGG..TAT	
CATTLE	CACAGTCTGA CTTCAGATAT ACTGTCTTTT TCTCTGGCTCC TTGGGCTTAG	
GOAT	AGCAGCCTGG CTTCAGATT. .CTGGCCTCT GCTT...T..	
HUMAN	AGCAGCCTGG CTTCAGATT. .CTGGCCTCT GCTTGGCT..	
CONSENSUS	TGCAGCCTGG CTTCAGATA. .CTGGCCTCT GCCTGGCTCC TTG.....A	
	-GCAGCCTGG CTTCAGATAT -CTGGCCTCT GCCTGGCTCC TTGGG--TA-	
RAT	232	MNF-1 NF-1 281
MOUSE	TTCCACCTTG TCCTTGCCCA GGTCCAAGAA AAAGCCCAGA ACCTTGGCAC	
CATTLE	GTCTACCTTG TCCTTGCCCA GGTCCAAGAA AAGGCCAGA ACCTTGGCAC	
GOAT	..CCACTTGT TGCTTTTCAA TGACCAAGAA .AAGCCCAGG CACTTGAAT	
HUMAN	..CCACTTGT TGCTTTTCAA TGACCAAGAA .AATCCCAGG CCCTTGAAT	
CONSENSUS	TCCCACCTTG CCCTTGTCAG TGACCAAGAA GAAGCCCAGC ACCTTGGCAC	
	TTCCACCTTG TCCTTGTCAA TGACCAAGAA AAAGCCCAG- ACCTTGGCAC	

	282	MYB		TCF-1	331
RAT	TGCTTTGCCA	GTTAATGTCT	AACCGAGGAA	TGTCTTGCTG	CCAAAAGGTG
MOUSE	TGTTTTGCCA	GTTAATGTCT	AACTGAGGAA	TGTCTTGCTG	CCAAAAGGTG
CATTLE	TTTTTACCCA	GTTAATTCT	AACTAAAGAA	CCTCTCGTTG	CCAAAAGATA
GOAT	TGTTTACTCA	GTTAATTCT	AACTAAAGAA	CCTCTTGTTG	CCAAAAGGTG
HUMAN	TGCTTTCCA	GTTAATTCT	AACTATGGAA	TCTCTTGCTG	TTAGAAGGTG
CONSENSUS	TGTTTCCA	GTTAATTCT	AACTAAGGAA	TCTCTTGCTG	CCAAAAGGTG
	332	FI-FII	GR	CF1	DEF 381
RAT	.CAAACAGAG	ACCTTGTATT	TCCAGGCACA	GGTGTGACCC	C CAATGTCA
MOUSE	.AAAACAGAG	ACCTTGTATT	TCCAGGCACA	GGTGTGACCC	C AATGTCA
CATTLE	TAACACAGAG	CCCTTGTAAAC	TCTGGGCACA	ACTGTGACCC	C AGTGTCA
GOAT	TAACACAGAG	CCCTTGTAGC	TGTGGGCACA	GCTGTGACCC	C CATGTCA
HUMAN	CGAAACAGTG	ACCTTGTATT	TCCGGGCACA	GCTGTGACCC	CCCATGTCA
CONSENSUS	TAACACAGAG	ACCTTGTATT	TCCGGGCACA	GCTGTGACCC	C CAATGTCA
	382	SBF-1	NF-AT3	GR	431
RAT	ATCATTITCT	GTCTCTAACT	ACCAGAGGAA	AAACTAACAA	CAACAGCCTC
MOUSE	ATCATTIT..T	GTGTCTAACT	CCCAGGGGAA	AAACTAACAA	CAACAGACTC
CATTLE	ATCATTIGGG	GTCTCTACCT	ATTAG	GGAA AA	GAACAA CAACCAACCTC
GOAT	ATCATTIGGG	GTCTCTACCT	ATTAG	GGAA AA	GAACAA CANCCACCTC
HUMAN	ATCATTIGGG	GTCTCTAGCT	ATTA	GGAA AA	AGAACAA CAACAACCTC
CONSENSUS	ATCATTIGGG	GTCTCTA-CT	ATTAG-	GGAA AAAAGAACAA	CAACAACCTC
	432	TCF-1	C-ETS-2		481
RAT	ATGGTTTGGG	AAAGGTGAAC	TCTATGCCAA	ATGGGAAGAAA	AAATTCTGAC
MOUSE	ATGGCTTGGG	AAAGGTGAAT	TCTATGCCAA	AAAGGGAGGA	AAGTTCT.AC
CATTLE	ACAGCCTAGA	AAAGGAAAAC	ACTGTGCAA	AAAGGGAAAAA	TATTCC..AC
GOAT	ACAGCCTANA	AAAGGAAAAC	ACTGTGCAA	AAAGGGAAAAA	TATTCC..AC
HUMAN	ACAGCTTGGG	CAAGGCCAAAC	ATTATGCCAG	AGGGAAAAAA	TATTCC..AC
CONSENSUS	ACAGCTTGGG	AAAGG-AAAC	ACTATGCCAA	AGGGAAAAAA	TATTCT-AC
	482		HSTF	HSF2	TFI1-I 531
RAT	CCCCCACAGAA	ACAATCTCAA	GAGGCAGAAC	CAGAGAATAA	TTGGAGG.GA
MOUSE	CCCCCACAGAA	ACAATCTCAG	AGGGCAGAAC	CAGAGAATAA	TCTGAGG.GA
CATTLE	CCCCATTAAT	ATA..ATTAA	AAACAGAAC	CAGAGGATCA	TTGGAGGAAA
GOAT	CCCCATTAAT	ATA..ATTAA	AAACAGAAC	CAGAGGATCA	TTGGAGGAGA
HUMAN	CCCCAAGAAA	ACAATATCAA	AAAACAGAAC	TAGAGACTAA	TTGGAGGAGA
CONSENSUS	CCCCA--AAA	ACAATATCAA	AAACAGAAC	CAGAGAATAA	TTGGAGGAGA
	532	CAC-BP	SP1	TFIID	581
RAT	GAGGGCCAGC	CAAGGGCAGA	CATA TATATA	TATATATTGA	TCACAGGCAC
MOUSE	GAGGGCCAGC	CAAGGGCAGG	CAAGTATATA	TTGA	TCACAGGCAC
CATTLE	GACTGCCAGT	GGGGGACAGA	TGTATATATA	TAGATATGAT	AGTCACCTAC
GOAT	GATTGCCAGT	GGGGGACAGA	TGTATATATA	TAGATATGAA	AGTCACCTAC
HUMAN	GATTGCCAGC	CTGGGGAAA	TGTGTATATA	TAAGTATGAG	GCACA.....
CONSENSUS	GA-TGCCAGC	C-GGGCAGA	TGTATATATA	TA-ATATGAA	-CACA---AC
	582	591			
RAT	TTACTTGTGA				
MOUSE	TTACTTGTGA				
CATTLE	TTGTAAAAGG				
GOAT	TTGTAAAAGG				
HUMAN	TCATCACCAAG				
CONSENSUS	TTAT-A--GG				

Sequence Identity of Uromodulin Promoters of Different Mammals

	Mouse	Rat	Goat	Cattle	Human
Mouse	N/C	90%	74%	73%	77%
Rat	90%	N/C	73%	74%	73%
Goat	74%	73%	N/C	94%	76%
Cattle	73%	74%	94%	N/C	78%
Human	77%	73%	76%	78%	N/C

Identity between sequence pairs are indicated as percentage identity. Note that promoters of two farm animals, goat and cattle, share 94% identity.

N/C: Sequences from the same species are 100% identical and are thus not compared.

Mouse versus rat **BestFit Results**

Refine

BESTFIT of: m.THP.promoter.log check: 5833 from: 1 to: 2092

REFORMAT of: m.THP.promoter.log.29746 check: 5833 from: 1 to: 2092
February 18, 2001 21:38
(No documentation)

to: rat589 check: 5372 from: 1 to: 581

Symbol comparison table: swgapdna.cmp CompCheck: 2335

Gap Weight: 50 Average Match: 10.000
Length Weight: 3 Average Mismatch: -9.000

Quality: 4177 Length: 585
Ratio: 7.315 Gaps: 8
Percent Similarity: 90.459 Percent Identity: 90.459

Match display thresholds for the alignment(s):

= IDENTITY
:= 5
:= 1

m.THP.promoter.log x rat589 September 22, 2004 10:34 ..

297 ccgaggaatgtcttgctgccaaaagggtgcaaacagagacaccttgttcc 346
1868 AGGCACAGGTGTGA.CCCCAATGTCAATCATT..TGTGTCTAACTCCCCA 1914
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
347 aggcacaggtgtgaccccaatgtcaatcatttctgtctctaactacca 396
1915 GGGGAAAAACTAACAAACACAGACTCATGGCTTGAAAGGTGAATTCTA 1964
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
397 gaggaaaaactaacaacaacagcctcatggttggaaaaggtgaactcta 446
1965 TGCCAAAAGGGAAGGAAAGTTCT.ACCCCCCACAGAAACAATCTCAGAGGG 2013
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
447 tgccaaatggaaagaaaaattctgacccccacagaaacaatctcaagagg 496
2014 CAGAACAGAGAATAATCTGAGGGAGAGGGCCAGCCAAGGGCAGGC.... 2059
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
497 cagaaggcagagaataattggagggagggccagccaagggcagacatat 546
2060 ..AAGTATATATTGATCACAGGCACTTACTTGTGA 2092
| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
547 atatatatatattgatcacaggcacttacttgtga 581

Input Sequence: m.THP.promoter.log

```
!!NA_SEQUENCE 1.0
REFORMAT of: m.THP.promoter.log.29746 check: 5833 from: 1
to: 2092 February 18, 2001 21:38
(No documentation)

m.THP.promoter.log Length: 2092 February 18, 2001 21:38
Type: N Check: 5833 ..

1 AATTTAAGAC TGGTGATTTC TTGAATTTC AGTGGGCTTG
```

[View Sequence](#)

Input Sequence: rat589

```
!!NA_SEQUENCE 1.0

rat589 Length: 581 September 18, 2001 10:45 Type: N
Check: 5372 ..

1 tagtcttgtc tgacagaggt ccagttgagg gatgtccaga
tggtcttgca

51 accgataact ttctcagaga ctctctctt cctgtctgga
ctcttagtggg
```

[View Sequence](#)

Mouse versus goat

BestFit Results

Refine

BESTFIT of: m.THP.promoter.log check: 5833 from: 1 to: 2092

REFORMAT of: m.THP.promoter.log.29746 check: 5833 from: 1 to: 2092
February 18, 2001 21:38
(No documentation)

to: g.THP.promoter18.log check: 4545 from: 1 to: 1630

REFORMAT of: g.THP.promoter18.log.22665 check: 4545 from: 1 to: 1630
February 18, 2001 21:37
(No documentation)

Symbol comparison table: swgapdna.cmp CompCheck: 2335

Gap Weight: 50 Average Match: 10.000
Length Weight: 3 Average Mismatch: -9.000

Quality: 1374 Length: 422
Ratio: 3.444 Gaps: 11
Percent Similarity: 74.300 Percent Identity: 73.791

Match display thresholds for the alignment(s):

= IDENTITY
:= 5
. = 1

m.THP.promoter.log x g.THP.promoter18.log September 22, 2004 10:32 ..

1677	AACATTCCCTTTATCCTAACACAGTCTGACTTCAGATATACTGTCTTTT	1726
1121	AGCACCCCTTCTCCTGGAGCAGCCTGGCTTCAGA.....T	1157
1727	CCTGGCTCCTTGGGCTTAGGTCTACCTTGCCCTGCCAGGTCCAAGAAA	1776
1158	TCTGGCCTCT...GCTTGGCTCCACTTGTGCTTTCAATGACCAAGAAA	1204
1777	AGGCCAGAACCTTGGCACTGTTTGCCAGTTAACGTCTAACTGAGGAAT	1826
1205	A.TCCCAGGCCCTTGGATTGTTACTCAGTTAACCTCTAACTAAAGAAC	1253
1827	GTCTTGCTGCCAAAAGGT.GAAAACAGAGACCTTGATTTCCAGGCACAG	1875
1254	CTCTTGTGCCAAAAGGTATAAACAGAGGCCCTGTAGCTGTGGGCACAG	1303
1876	GTGTGACCCCAATGTCAATCATTT..TGTGTCTAACCTCCAGGGAAAAA	1923
1304	CTGTGACCCCCATGTCAATCATTTGGGTCTTACCTATTAGGG...AAA	1350
1924	CTAACACAACAGACTCATGGCTTGGAAAAGGTGAATTCTATGCCAAAAG	1973
1351	AGAACAAACANCCACCTCACAGCTTANAAAAGGAAAACACTGTGTCAAAAG	1400

```

1974 GGAAGGAAAGTTCTACCCCCACAGAAACAATCTCAGAGGGCAGAAGCAGA 2023
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |
1401 GGAA.AAATATTCCACCCCCATTAAAATAAT.TAACAGAACCGAGA 1447

2024 GAATAATCTGAGG.GAGAGGGCCAGCCAAGGGCAG..GCAAGTATATATT 2070
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |
1448 GGATCATTGGAGGAGAGATTGCCAGTGGGGACAGATGTATATATAGA 1497

2071 GATCACAGGCACTTACTTGTGA 2092
||| ||| ||| ||| ||| ||| |
1498 TATGAAAGTCACCTACTTGTAA 1519

```

Input Sequence: m.THP.promoter.log

```

!!NA_SEQUENCE 1.0
REFORMAT of: m.THP.promoter.log.29746 check: 5833 from: 1
to: 2092 February 18, 2001 21:38

(No documentation)

m.THP.promoter.log Length: 2092 February 18, 2001 21:38
Type: N Check: 5833 ..

1 AATTTAAGAC TGGTTGATT CTTGAATTTC AGTGGGCTTG

```

[View Sequence](#)

Input Sequence: g.THP.promoter18.log

```

!!NA_SEQUENCE 1.0
REFORMAT of: g.THP.promoter18.log.22665 check: 4545 from:
1 to: 1630 February 18, 2001 21:37

(No documentation)

g.THP.promoter18.log Length: 1630 February 18, 2001 21:37
Type: N Check: 4545 ..

1 ACTATAGGGC ACCCGTGGTC GACGGCCCGG GCTGGTAAAG

```

[View Sequence](#)

Mouse versus cattle

BestFit Results

Refine

BESTFIT of: m.THP.promoter.log check: 5833 from: 1 to: 2092

REFORMAT of: m.THP.promoter.log.29746 check: 5833 from: 1 to: 2092
 February 18, 2001 21:38
 (No documentation)

to: cattle.THP.promoter.txt check: 7177 from: 1 to: 626

REFORMAT of: cattle.THP.promoter.txt.27851 check: 7177 from: 1 to: 626
 February 18, 2001 21:37
 (No documentation)

Symbol comparison table: swgapdna.cmp CompCheck: 2335

Gap Weight: 50 Average Match: 10.000
 Length Weight: 3 Average Mismatch: -9.000

Quality: 1284 Length: 439
 Ratio: 3.109 Gaps: 12
 Percent Similarity: 72.973 Percent Identity: 72.973

Match display thresholds for the alignment(s):

| = IDENTITY
 : = 5
 . = 1

m.THP.promoter.log x cattle.THP.promoter.txt September 22, 2004 10:26 ..

```

1660 TTCCAGTCTCAAAGCAACATTCTTTATCCTAACACAGTCTGACTTC 1709
| | |||| | | | | | | | | | | | | | | | | | | | | | |
168 tcccaagtgggacagtgagcacccctttctcctggagcagcctggcttc 217

1710 AGATATACTGTCTTTCTGGCTCCTGGGCTTAGGTCTACCTTGCCT 1759
| | | | | | | | | | | | | | | | | | | | | | | | | |
218 aga.....ttctggcctct...gctt...tccacttttgct 248

1760 TGCCCAGGTCCAAGAAAAGGCCAGAACCTGGCACTGTTTGCCAGTTA 1809
| | | | | | | | | | | | | | | | | | | | | | | | | |
249 ttcaatgaccaagaaaa.gcccaggcaatttgcatacccaatgtt 297

1810 ATGTCTAACTGAGGAATGTCTGCTGCCAAAAGGT.GAAAACAGAGACCT 1858
| | | | | | | | | | | | | | | | | | | | | | | | | |
298 atttctaactaaagaacctctcggtccaaaagatataaaacagagccct 347

1859 TGTATTTCCAGGGCACAGGTGTGACCCCAATGTCAATCATTT..TGTGTCT 1906
| | | | | | | | | | | | | | | | | | | | | | | | | |
348 tgtaactctgggcacaactgtgaccccaatgtcaatcattgggtctct 397

1907 AACTCCCAGGGAAAAACTAACAAACAACAGACTCATGGCTTGGAAAAGGT 1956
| | | | | | | | | | | | | | | | | | | | | | | | | |
398 acctatttaggg...aaaagaacaacaaccacacctcacagcctagaaaagga 444

```

1957 GAATTCTATGCCAAAAGGGAAAGGAAAGTTCTACCCCCACAGAAACAATCT 2006
 ||| ||||| ||||| ||||| ||||| |||||
 445 aaacactgtgtcaaaaggaa.aaatattccaccccccattaaaataat.t 492

2007 CAGAGGGCAGAAGCAGAGAATAATCTGAGG.GAGAGGGCCAGCCAAGGGC 2055
 ||| ||||| ||||| ||||| ||||| |||||
 493 aaga.aacagaaccagaggatcatggagggaaagactgccagtggggac 541

2056 AG..GCAAGTATATATTGATCACAGGCACTTACTTGTGA 2092
 ||| ||| ||||| ||||| ||||| |||||
 542 agatgtatatatagatatgatagtcacctacttgtaa 580

Input Sequence: m.THP.promoter.log

```
! !NA_SEQUENCE 1.0
REFORMAT of: m.THP.promoter.log.29746 check: 5833 from: 1
to: 2092 February 18, 2001 21:38

(No documentation)

m.THP.promoter.log Length: 2092 February 18, 2001 21:38
Type: N Check: 5833 ..

1 AATTAAAGAC TGGTGATT CTTGAATTTC AGTGGGCTTG
```

[View Sequence](#)

Input Sequence: cattle.THP.promoter.txt

```
! !NA_SEQUENCE 1.0
REFORMAT of: cattle.THP.promoter.txt.27851 check: 7177
from: 1 to: 626 February 18, 2001 21:37

(No documentation)

cattle.THP.promoter.txt Length: 626 February 18, 2001
21:37 Type: N Check: 7177 ..

1 aatttcttga ttcacagagc atctggtcca atgatgtctg
```

[View Sequence](#)

BestFit Results

Mouse versus human

Refine

BESTFIT of: m.THP.promoter.log check: 5833 from: 1 to: 2092

REFORMAT of: m.THP.promoter.log.29746 check: 5833 from: 1 to: 2092
February 18, 2001 21:38
(No documentation)

to: human.THP.promoter.txt check: 7451 from: 1 to: 620

REFORMAT of: human.THP.promoter.txt.30475 check: 7451 from: 1 to: 620
February 18, 2001 21:36
(No documentation)

Symbol comparison table: [swgapdna.cmp](#) CompCheck: 2335

Gap Weight: 50 Average Match: 10.000
 Length Weight: 3 Average Mismatch: -9.000
 Quality: 1790 Length: 407
 Ratio: 4.509 Gaps: 7
 Percent Similarity: 76.590 Percent Identity: 76.590

```
Match display thresholds for the alignment(s):
    | = IDENTITY
    : = 5
    . = 1
```

m.THP.promoter.log x human.THP.promoter.txt September 22, 2004 10:33 ..

1682	TCCTTTATCCTAACACAGTCTGACTTCAGATATACTGTCTTTCCCTGG	1731
173	tcctttgtccttatgcagcctggcttcag..atactggctctgcctgg	220
1732	CTCCTGGGCTTAGGTCTACCTTGTCCCTGCCAGGTCCAAGAAAAGGCC	1781
221	ctccttgatc.....ccaccctgcccattgtcagtgaccaagaagaagcc	264
1782	CAGAACCTGGCACTGTTGCCAGTTAATGTCTAACTGAGGAATGTCTT	1831
265	cagcaccttggcactgcttccagttaaattctaactatggaatcttctt	314
1832	GCTGCCAAAAGGTG . AAAACAGAGACCTTGATTTCCAGGCACAGGTGTG	1880
315	gctgttagaagggtgcgaaacagtgaccttgtatttccggcacaggtgtg	364
1881	A .. CCCCAATGTCAATCATTTGTGTCTAACTCCCAGGGAAAAACTAAC	1928
365	accccccaatgtcaatcatttgggtctctagctatttagaaaaaa.gaac	413
1929	AACAACAGACTCATGGCTTGGAAAAGGTGAATTCTATGCCAAAAGGGAAG	1978
414	aacaacaacacctcacagcttggacaaggcaaacattatqcccaqqa.qqaaa	462

1979 GAAAGTTCTACCCCCACAGAAACAATCTCAGAGGGCAGAAGCAGAGATA 2028
|| ||||| ||||| ||||| ||||| ||||| ||||| |||||
463 aaatattccaccccaagaaaacaatatcaaaaaacagaactagagacta 512

2029 ATCTGAGG.GAGAGGGCCAGCCAAGGGCAGGCAAGTATATATTGATCACA 2077
|| ||||| ||||| ||||| ||||| ||||| ||||| |||||
513 attggaggagagattgccagcctgggcaaatgttatataagtatga 562

2078 GGCAC TT 2084
||||| |
563 ggcacat 569

Input Sequence: m.THP.promoter.log

```
!INA_SEQUENCE 1.0
REFORMAT of: m.THP.promoter.log.29746 check: 5833 from: 1
to: 2092 February 18, 2001 21:38

(No documentation)

m.THP.promoter.log Length: 2092 February 18, 2001 21:38
Type: N Check: 5833 ..

1 AATTTAAGAC TGGTTGATTT CTTGAATTTC AGTGGGCTTG
```

[View Sequence](#)

Input Sequence: human.THP.promoter.txt

```
!INA_SEQUENCE 1.0
REFORMAT of: human.THP.promoter.txt.30475 check: 7451
from: 1 to: 620 February 18, 2001 21:36

(No documentation)

human.THP.promoter.txt Length: 620 February 18, 2001 21:36
Type: N Check: 7451 ..

1 cagagtgggt caggtccagt gatgtctgaa ctaccttctg
```

[View Sequence](#)

BestFit Results

Rat versus goat

Refined

BESTFIT of: Rat.THP.promoter.txt check: 1164 from: 1 to: 625

REFORMAT of: Rat.THP.promoter.txt.27903 check: 1164 from: 1 to: 625
February 18, 2001 21:37
(No documentation)

to: g.THP.promoter18.log check: 4545 from: 1 to: 1630

REFORMAT of: g.THP.promoter18.log.22665 check: 4545 from: 1 to: 1630
February 18, 2001 21:37
(No documentation)

Symbol comparison table: swgapdna.cmp CompCheck: 2335

Gap Weight: 50 Average Match: 10.000
Length Weight: 3 Average Mismatch: -9.000

Quality: 1640 Length: 462
Ratio: 3.744 Gaps: 9
Percent Similarity: 73.733 Percent Identity: 73.272

Match display thresholds for the alignment(s) :

| = IDENTITY
: = 5
. = 1

Rat.THP.promoter.txt x g.THP.promoter18.log September 22, 2004 11:11

168 ctgttatctaacccaggctggcttcagatattgtcttttccgc 217
|| || || || || || || || || | | | | |
1128 CTTTTCTCCTGGAGCAGCCTGGCTTCAGA TTCTGGCC 1165

268 cttggcactgcttgcagttaatgtctaacccgaggaatgtcttgctgcc 317
||| | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
1215 CTTGGCAGTTCGTTTACTTGATTTGATTTGATTTGATTTGATTTG

417 cagcctcatggttggaaaaggtaacttatgccaaatggaaagaaaaa 466
| | | | | | : | | | | | | | | | | | | | | | | | | | | |
1361 CCACCTCACAGCCTANAAAAGGAAAACACTGTGTCAAAAGGGAAAAATAT 1410

467 ttctgaccccacagaaacaatctcaagaggcagaagcagagaataattg 516
|| | || || || | || | || | || | || | || | || |
472 tcc..accccattaaaataa..ttaagaaacagaaccagaggatcattg 517

517 gagggagagggccagccaagggcagacatatatatatatattgtatcac 566
|| | || | || || | || | || | || | || | || | || |
518 gag...gaaagactgccagtggggacagatgtatatatagatatgat 564

567 aggcaacttacttgtaatggaccagtcc...gtcctgggttcaggtaag 613
|| | || | || || | || | || | || | || | || | || |
565 agtcacaccttgcataaggattaatttaccccttctggttcaggtaag 614

614 actgtctggag 624
|| | || | ||
615 gctatctgcag 625

Input Sequence: Rat.THP.promoter.txt

```
!!NA_SEQUENCE 1.0
REFORMAT of: Rat.THP.promoter.txt.27903 check: 1164 from:
1 to: 625 February 18, 2001 21:37
(No documentation)

Rat.THP.promoter.txt Length: 625 February 18, 2001 21:37
Type: N Check: 1164 ..
1 ctagtcttgt ctgacagagg tccagtttag ggatgtccag
```

[View Sequence](#)

Input Sequence: cattle.THP.promoter.txt

```
!!NA_SEQUENCE 1.0
REFORMAT of: cattle.THP.promoter.txt.27851 check: 7177
from: 1 to: 626 February 18, 2001 21:37
(No documentation)

cattle.THP.promoter.txt Length: 626 February 18, 2001
21:37 Type: N Check: 7177 ..
1 aatttcttga ttcacagagc atctggcca atgatgtctg
```

[View Sequence](#)

Rat versus Human

BestFit Results

Refine

BESTFIT of: Rat.THP.promoter.txt check: 1164 from: 1 to: 625

REFORMAT of: Rat.THP.promoter.txt.27903 check: 1164 from: 1 to: 625
February 18, 2001 21:37
(No documentation)

to: human.THP.promoter.txt check: 7451 from: 1 to: 620

REFORMAT of: human.THP.promoter.txt.30475 check: 7451 from: 1 to: 620
February 18, 2001 21:36
(No documentation)

Symbol comparison table: swgapdna.cmp CompCheck: 2335

Gap Weight: 50 Average Match: 10.000
Length Weight: 3 Average Mismatch: -9.000

Quality: 2040 Length: 588
Ratio: 3.611 Gaps: 10

Percent Similarity: 73.297 Percent Identity: 73.297

Match display thresholds for the alignment(s):

| = IDENTITY
: = 5
. = 1

Rat.THP.promoter.txt x human.THP.promoter.txt September 22, 2004 11:14 ...

```

340 gtatttccaggcacaggtgtga.ccccaatgtcaatcatttctgtctc 388
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |
344 gtatttccgggcacaggtgtgacccccataatgtcaatcatttgggtctc 393
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |
389 taactaccagagaaaaactaacaacaacaacgcctcatggtttggaaaagg 438
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |
394 tagctatta...ggaaaaagaacaacaacaacctcacagcttggacaagg 440
439 tgaactctatgccaaatggaaagaaaaattctgacccccacagaaaaaat 488
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |
441 caaacattatgccaggaggaaaaatattcc..acccccaagaaaaacaat 488
489 ctcagaaggcagaaggcagaataattggagg..gagaggggccagccaagg 537
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |
489 atcaaaaaacagaactagagactaattggaggagattgccagcctggg 538
538 gcagacatatatatatatattgatcacaggcacttacttgtgaatgg 587
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |
539 gcaaattgttatataagtat.....gaggcacatcatcaccagacta 582
588 ccagtctgtcctgggttcaggtaagactgtctggagc 625
| | | | | | | | | | | | | | | | | | | | | | | |
583 actctaccttctggcttcaggtaaggctatctgttagc 620

```

Input Sequence: Rat.THP.promoter.txt

```

!!NA_SEQUENCE 1.0
REFORMAT of: Rat.THP.promoter.txt.27903 check: 1164 from:
1 to: 625 February 18, 2001 21:37
(No documentation)

Rat.THP.promoter.txt Length: 625 February 18, 2001 21:37
Type: N Check: 1164 ...
1 ctagtcttgt ctgacagagg tccagttgag ggatgtccag

```

[View Sequence](#)

Input Sequence: human.THP.promoter.txt

```
! !NA_SEQUENCE 1.0
REFORMAT of: human.THP.promoter.txt.30475 check: 7451
from: 1 to: 620 February 18, 2001 21:36

(No documentation)

human.THP.promoter.txt Length: 620 February 18, 2001 21:36
Type: N Check: 7451 ..

1 cagagtgggt caggtccagt gatgtctgaa ctaccttctg
```

[View Sequence](#)

goat versus cattle

BestFit Results

Refine

BESTFIT of: g.THP.promoter18.log check: 4545 from: 1 to: 1630

REFORMAT of: g.THP.promoter18.log.22665 check: 4545 from: 1 to: 1630
February 18, 2001 21:37
(No documentation)

to: cattle.THP.promoter.txt check: 7177 from: 1 to: 626

REFORMAT of: cattle.THP.promoter.txt.27851 check: 7177 from: 1 to: 626
February 18, 2001 21:37
(No documentation)

Symbol comparison table: swgapdna.cmp CompCheck: 2335

Gap Weight: 50 Average Match: 10.000
Length Weight: 3 Average Mismatch: -9.000

Quality: 5545 Length: 628
Ratio: 8.872 Gaps: 1

Percent Similarity: 94.560 Percent Identity: 94.240

Match display thresholds for the alignment(s):
| = IDENTITY
: = 5
. = 1

g.THP.promoter18.log x cattle.THP.promoter.txt September 22, 2004 10:36 ..

937 AATTCTTGATTCACAGAGCATCTGGTCCAATGATGTCTGAATTATCTGC 986

||||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

1 aattcttgattcacagagcatctggtccaatgatgtctgaattgccttc 50

987 TGTCTCTGACCTTCAGCCATTCTCAGCTCCTTCCTGATCACATTGGAC 1036

||||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

51 tgtctctgaccttcaggcattctcagtccttcctgctcacatcgggac 100

1037 CCCAGGGGAGCTGGCTGAATCTGTGAGGATGGCATTGCTTGGAAATTAA 1086

||||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

101 cccagggaaagctggtaacccatgaggatggaaacttgcttggaaactga 150

1087 GTGCCACAAAGTACACATCCTGGGGACATGAGCACCCCTTTCTCC 1136

||||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

151 gtggccacaagtatacatcccagtggggacagttagcacccctttctcc 200

1137 TGGAGCAGCCTGGCTTCAGATTCTGGCCTCTGCTTGGCTCCACTTGTGC 1186

||||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

201 tggagcagcctggcttcagattctggcctctgctt...tccactttgtgc 247

1187 TTTTCAATGACCAAGAAAATCCCAGGCCCTTGGAAATTGTAACTCAGTTA 1236

||||||| ||||| ||||| ||||| ||||| ||||| ||||| |||||

248 ttttcaatgaccaagaaaagcccaggcacttgaattttaccagtt 297

1237 ATTTCTAACTAAAGAACCTCTTGTGCCAAAAGGTATAAACAGAGCCCT 1286
 |||||
 298 atttctaactaaagaacctctcggtccaaaagatataaaacagagccct 347

1287 TGTAGCTGTGGGCACAGCTGTGACCCCCATGTCAATCATTGGGTCTCT 1336
 |||||
 348 tgtaactctggcacaactgtgaccccagtgtcaatcattgggtctct 397

1337 ACCTATTAGGGAAAAGAACACACANCCACCTCACAGCCTANAAAAGGAAAA 1386
 |||||:|||||:|||||:|||||:|||||
 398 acctattagggaaaagaacaacaaccacacctcacagcctagaaaaaggaaaa 447

1387 CACTGTGTCAAAAGGAAAAATATTCCACCCCCATTAAAATAATTAGAA 1436
 |||||
 448 cactgtgtcaaaaggaaaaatattccaccccatthaataattaagaa 497

1437 ACAGAACCCAGAGGATCATGGAGGAGATTGCCAGTGGGGACAGATGT 1486
 |||||
 498 acagaaccagaggatcatggagaaagactgccagtggggacagatgt 547

1487 ATATATATAGATATGAAAGTCACCTACTTGTAAAAGGATTAATTCTACCT 1536
 |||||
 548 atatatatagatatgatagtcacctacttgtaaaaggattaattctacct 597

1537 TTCTGGTTTCAGGTAAGGCTATCTGCAG 1564
 |||||
 598 ttctggtttcaggttaaggctatctgcag 625

Input Sequence: g.THP.promoter18.log

```
!!NA_SEQUENCE 1.0
REFORMAT of: g.THP.promoter18.log.22665 check: 4545 from:
1 to: 1630 February 18, 2001 21:37
(No documentation)

g.THP.promoter18.log Length: 1630 February 18, 2001 21:37
Type: N Check: 4545 ..

1 ACTATAGGGC ACGCGTGGTC GACGGCCCGG GCTGGTAAAG
```

[View Sequence](#)

Input Sequence: cattle.THP.promoter.txt

```
!!NA_SEQUENCE 1.0
REFORMAT of: cattle.THP.promoter.txt.27851 check: 7177
from: 1 to: 626 February 18, 2001 21:37

(No documentation)

cattle.THP.promoter.txt Length: 626 February 18, 2001
21:37 Type: N Check: 7177 ..

1 aatttcttga ttcacagago atctggtcca atgatgtctg
```

[View Sequence](#)

BestFit Results

Refine

BESTFIT of: q.THP.promoter18.log check: 4545 from: 1 to: 1630

REFORMAT of: g.THP.promoter18.log.22665 check: 4545 from: 1 to: 1630
February 18, 2001 21:37
(No documentation)

to: human.THP.promoter.txt check: 7451 from: 1 to: 620

Symbol comparison table: swqapdna.cmp CompCheck: 2335

Gap Weight: 50 Average Match: 10.000
Length Weight: 3 Average Mismatch: -9.000

Quality: 2836 Length: 633
Ratio: 4.596 Gaps: 8
Percent Similarity: 76.325 Percent Identity: 75.993

```
Match display thresholds for the alignment(s):
    | = IDENTITY
    : =      5
    . =      1
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g.THP.promoter18.log x human.THP.promoter.txt September 22, 2004 11:22

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999 TCAGGCCATTCTCAGCTCCCTTCCTGATCACATTGGGACCCCAGGGGAGCT 1048
||||| ||||| ||||| || | | | | | | | | | | | |
51 tcaagccattctcagctccctcttactttatctggattcta a 93

```

1049 GGCTGAATCTGTGAGGATGGCATTGCTTGAAATTAAAGTGGCC...ACA 1095
      |||||    |||||  |||||  ||| |  | | | | | | | | | | | |
  94 ggctgatctcatqqaatqqqtqttcaqaaggataccctctccaagaca 143

```

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| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
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1237 ATTTCTAACTAAAGAACCTCTTGTGCCAAAAGGTATAAAACAGAGCCCT 1286
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 293 atttctaaactatggaatctcttgctgttagaaggtgcgaaacagtgcacct 342

1287 TGTAGCTGTGGGCACAGCTGTGACCCCC..ATGTCAATCATTGGGGTCT 1334
 ||||| | ||||| ||||| ||||| ||||| |||||
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 ||||| ||||| ||||| ||||| :| ||||| ||||| :| |||||
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1385 AACACTGTGTCAAAAGGGAAAAAATTCCACCCCCATTAAAATA..ATTA 1432
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
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1433 AGAAACAGAACCAAGAGGATCATTGGAGGAGAGATTGCCAGTGGGGACAG 1482
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
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1533 ACCTTTCTGGTTTCAGGTAAGGCTATCTGCAGC 1565
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(No documentation)

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[View Sequence](#)

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from: 1 to: 620 February 18, 2001 21:36
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(No documentation)

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Type: N Check: 7451 ..
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[View Sequence](#)

BestFit Results

Cattle versus human

Refine

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February 18, 2001 21:37
(No documentation)

to: human.THP.promoter.txt check: 7451 from: 1 to: 620

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February 18, 2001 21:36
(No documentation)

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Length Weight: 3 Average Mismatch: -9.000

Quality: 2816 Length: 637
Ratio: 4.594 Gaps: 10

Percent Similarity: 77.815 Percent Identity: 77.815

Match display thresholds for the alignment(s):

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cattle.THP.promoter.txt x human.THP.promoter.txt September 22, 2004 11:20 ..

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14 cagagtgggtcaggtccagtgtctgaactacacctctggttctgactt 50

63 tcaggcattctcagtccttctgtcacatgggaccggagggact 112

||||| ||||||| || || || || || || || || || || || || || ||
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113 ggttgaacccatgaggatgaaacttgcggaaactgagtgcc..... 156

|| || | || || || || || || || || || || || || || || || ||
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|| || | || || || || || || || || || || || || || || || ||
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Input Sequence: cattle.THP.promoter.txt

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from: 1  to: 626  February 18, 2001 21:37

(No documentation)

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21:37  Type: N  Check: 7177  ..

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Input Sequence: human.THP.promoter.txt

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from: 1 to: 620 February 18, 2001 21:36
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(No documentation)

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Type: N Check: 7451 ..
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1 cagagtgggt caggtccagt gatgtctgaa ctaccttctg
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[View Sequence](#)

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Tamm-Horsfall Glycoprotein: Biology and Clinical Relevance

Franca Serafini-Cessi, MD, Nadia Malagolini, PhD, and Daniela Cavallone, PhD

• Tamm-Horsfall glycoprotein (THP) is the most abundant urinary protein in mammals. Urinary excretion occurs by proteolytic cleavage of the large ectodomain of the glycosyl phosphatidylinositol-anchored counterpart exposed at the luminal cell surface of the thick ascending limb of Henle's loop. We describe the physical-chemical structure of human THP and its biosynthesis and interaction with other proteins and leukocytes. The clinical relevance of THP reported here includes: (1) involvement in the pathogenesis of cast nephropathy, urolithiasis, and tubulointerstitial nephritis; (2) abnormalities in urinary excretion in renal diseases; and (3) the recent finding that familial juvenile hyperuricemic nephropathy and autosomal dominant medullary cystic kidney disease 2 arise from mutations of the THP gene. We critically examine the literature on the physiological role and mechanism(s) that promote urinary excretion of THP. Some lines of research deal with the in vitro immunoregulatory activity of THP, termed uromodulin when isolated from urine of pregnant women. However, an immunoregulatory function in vivo has not yet been established. In the most recent literature, there is renewed interest in the capacity of urinary THP to compete efficiently with urothelial cell receptors, such as uroplakins, in adhering to type 1 fimbriated *Escherichia coli*. This property supports the notion that abundant THP excretion in urine is promoted in the host by selective pressure to obtain an efficient defense against urinary tract infections caused by uropathogenic bacteria. *Am J Kidney Dis* 42: 658-676.

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INDEX WORDS: Tamm-Horsfall protein (THP); uromodulin; urinary proteins; glycosyl phosphatidylinositol (GPI)-anchored proteins; cast nephropathy; myeloma nephropathy; stone disease; interstitial nephritis; *Escherichia coli*; familial juvenile hyperuricemic nephropathy (FJHN); medullary cystic kidney disease 2 (MCKD2).

IN 1950, IGOR TAMM and Frank Horsfall¹ used a salt-precipitation procedure to isolate a potent inhibitor of viral hemagglutination from urine of healthy individuals. Subsequently, the same investigators² undertook the chemical characterization of mucoprotein and the way in which it inhibits hemagglutination induced by influenza, mumps, and Newcastle disease viruses. In general terms, the 2 investigators sought to obtain evidence of the (then) putative enzymatic activity associated with viruses and identify inhibitors preventing viruses from binding to susceptible cells. They reported that neuraminidase treatment abolished the inhibitory effect in the viral hemagglutination assay. This observa-

tion persuaded Gottschalk³ and Odin⁴ to analyze the carbohydrate moiety of urinary mucoprotein. Both studies established that carbohydrate content accounts for more than 20% to 25% of mucoprotein weight, and sialic acid is abundantly present. Urinary mucoprotein has since become known as Tamm-Horsfall glycoprotein or Tamm-Horsfall protein (THP).

In 1964, Bayer⁵ used electron microscopy to confirm the binding of influenza virus to urinary mucoprotein. Urinary THP was visualized as a network of filaments composed of smaller fibrils with a diameter of 4 to 24 nm, but the length could not be detected because of their tendency to form end-to-end aggregates. In the 1970s, Albert Neuberger's laboratory in London showed that THP: (1) migrates in sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) as a single molecular unit with an apparent molecular weight of 80 to 90 kd; (2) is the most abundant protein in normal urine, excreted at the rate of approximately 50 mg/d; and (3) is present in urine of other mammals.⁶⁻⁹

THP had been constantly under the attention of investigators working in the field of glycoproteins or nephrologists and urologists (according to PubMed from January 1967 to January 1985, almost 200 articles concerning Tamm-Horsfall glycoprotein were published) when, in 1985, it was rediscovered by Muchmore and Decker.¹⁰

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doi:10.1053/S0272-6386(03)00829-1

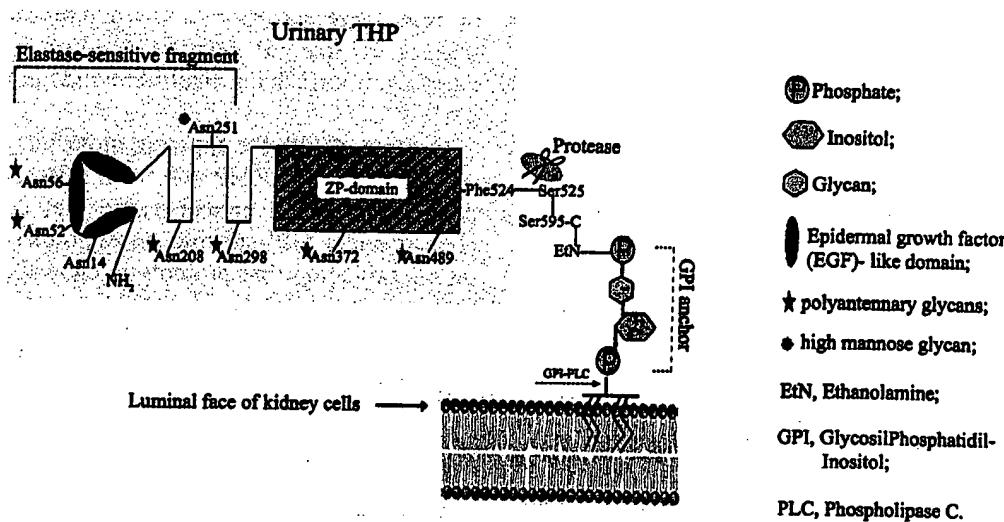


Fig 1. Structural model of urinary THP (yellow area) and its renal GPI-anchored counterpart.

These investigators isolated a glycoprotein from urine of pregnant women by a method that includes lectin-affinity chromatography. They named the protein uromodulin because in some conditions, it showed in vitro immunosuppressive activity. A similar property previously was shown to be present in THP.^{11,12} In 1987, the complementary DNA (cDNA) of uromodulin was cloned, and its identity with the cDNA of human THP was shown.^{13,14}

PHYSICAL-CHEMICAL STRUCTURE OF URINARY THP AND ITS RENAL GLYCOSYL PHOSPHATIDYLINOSITOL-ANCHORED COUNTERPART

The primary structure of human THP has been predicted by sequencing the cDNA.^{13,14} On the

basis of the *N*-terminal sequence (Asp-Thr-Ser-Glu-Ala) found in urinary THP, Pennica et al¹³ assigned 616 amino acids to THP, in that the first 24 *N*-terminal amino acids predicted by cDNA represent the signal peptide lacking in the mature protein (we have used amino acid-mapping according to Pennica et al,¹³ to indicate both the *N*-glycosylation sites and all cited amino acids throughout the text). The protein includes 48 cysteine residues, 8 potential *N*-glycosylation sites, 3 epidermal growth factor domains, 1 zona pellucida (ZP)-like domain, and, at the C-terminus, 1 stretch of hydrophobic amino acids, similar to that of proteins that acquire a glycosyl phosphatidylinositol (GPI) anchor. This hydrophobic sequence acts as a signal for endoplasmic reticulum (ER)-transpeptidase, which, after cleav-

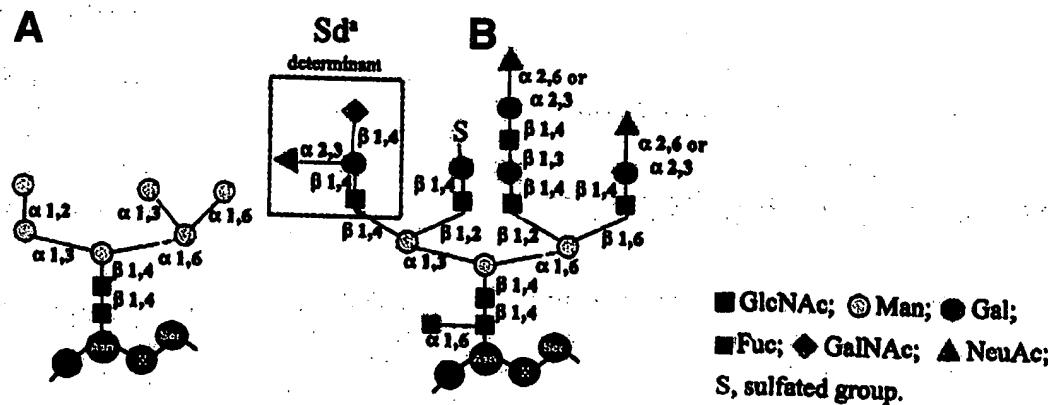


Fig 2. Representative structures of human THP *N*-glycans: (A) the major high-mannose sequence⁴⁷ and (B) the structure of a tetra-antennary chain showing the various nonreducing terminal units.^{38,39}

age of the hydrophobic peptide, adds the pre-formed GPI anchor to the "new" C-terminus (Fig 1).¹⁵ Rindler et al¹⁶ transfected THP cDNA into HeLa cells and showed that THP is a GPI-anchored protein. Fatty acids bound to the GPI anchor allow insertion of THP into the outer leaflet of the plasma membrane (Fig 1). DNA encoding THP in the rat, mouse, and calf also was sequenced,¹⁷⁻¹⁹ and the THP gene was reported present in all vertebrate classes.²⁰ The human THP gene was assigned to chromosome 16p12.3-16p13.11.²¹ The mouse THP gene promoter was isolated, and by using the transgenic mouse approach, a sequence of 3.0 kb was shown to be sufficient to drive kidney expression of a heterologous reporter gene.²²

On the basis of their amino acid sequence, a new class of GPI-anchored proteins, broadly homologous to THP, was identified. This class includes pancreatic glycoprotein 2 (GP2), which shows a 53% identity and 85% similarity to the human THP amino acid sequence^{17,23}; a sea urchin sperm membrane protein²⁴; and ZP2- and ZP3-glycoproteins, which participate in forming the transparent coat (zona pellucida) surrounding the eggs of all placental mammals.²⁵ GP2 is the major protein present in the zymogen secretory granule membrane of the pancreas and very likely is involved in the regulated release of zymogens.²⁶ Recently, Jovine et al²⁷ showed that (1) the peptide encompassing amino acids 1 to 291 of human THP is selectively degraded by pancreatic elastase, and (2) the residual C-terminal peptide chain of approximately 48 kd that THP shares with ZP2- and ZP3-glycoproteins (ZP domain) is responsible for polymerization of all these glycoproteins (Fig 1).

The first electron microscope and ultracentrifuge studies attributed to urinary THP an axial ratio of 250 and a molecular weight of $7 \times 10^6 M_r$.^{28,29} Subsequent studies showed that THP consists of protofilaments with a double-helix structure that, in polymer form, acquire a ribbon-shaped structure 1,500 to 4,000 nm in length and are organized in a large network.^{5,27} In SDS-PAGE, THP shows an apparent molecular weight of 80 kd in nonreducing conditions or 97 kd when the glycoprotein is reduced with 2-mercaptoethanol.^{7,30} The decrease in apparent molecular weight in nonreducing conditions depends on a high degree of polypeptide constraint imposed

by the large number of intrachain disulfide bridges, in that all cysteine residues are converted into intrachain disulfide bonds in urinary THP.³¹

Compared with the apparent molecular weight of human urinary THP, the renal GPI-anchored counterpart has a greater value that does not change under GPI-specific phospholipase treatment, indicating that proteolytic cleavage in the juxtamembrane region of the THP ectodomain is responsible for the urinary excretion that occurs.³⁰ Consistent with this, recombinant THP expressed by HeLa cells is released mainly devoid of ethanolamine, the residue responsible for binding the GPI anchor to the THP C-terminus³² (Fig 1). According to Fukuoka and Kobayashi,³³ proteolytic cleavage occurs between amino acids 524 (phenylalanine) and 525 (serine), ie, 66 amino acids upstream of the putative C-terminal residue bound to the GPI anchor.

THE GLYCOMOIEITY STRUCTURE OF THP

Carbohydrates account for approximately 30% of the weight of human THP and consist mainly of *N*-linked glycans of di-, tri-, and tetra-antennary type^{6,34-36}; however, 1 *N*-glycosylation site carries high mannose sequences.³⁷ The tetra-antennary chains are elongated in a sizable percentage by repeating *N*-acetyllactosamine sequences and show marked heterogeneity because of differences in the extent of sialylation, fucosylation, and sulfation (Fig 2).^{38,39} Of the 8 potential *N*-glycosylation sites predicted by amino-acid sequence, only 7 are actually glycosylated.⁴⁰

In addition, *N*-acetylgalactosamine (GalNAc) in β 1,4 linkage to galactose occurs in α 2,3 sialylated glycans, producing the GalNAc β 1,4(NeuAco2,3)Gal β 1,4GlcNAc-sequence known as Sd^a antigen (Fig 2).⁴¹ This antigen was originally described as a blood group determinant inherited as a dominant characteristic present in approximately 90% of the Caucasian population.⁴² The GalNAc residue is the immunodominant sugar, confirmed because THP from Sd^a-positive individuals contains 1% to 2% of GalNAc, whereas THP from Sd^a-negative individuals does not contain GalNAc.⁴³ Biosynthesis of Sd^a determinant depends on a specific β -GalNAc-transferase (Sd^a- β GalNAc-transferase) first described in our laboratory.^{44,45} Sd^a- β GalNAc-transferase activity is detected easily in the outer

medulla of human kidney, but not in the cortex, suggesting that the Sd^a antigen is carried prevalently by THP synthesized by cells from the thick ascending limb (TAL) of Henle's loop, rather than those from the early region of convoluted distal tubules.⁴⁶

In human THP, high-mannose glycans are carried by Asn₂₅₁,⁴⁰ and Man₆GlcNAc₂ is the preponderant structure over Man₇GlcNAc₂ and Man₅GlcNAc₂.^{37,47} Recombinant THP expressed by transfected HeLa cells also bears high-mannose sequences similar to those found in urinary THP, indicating that the occurrence of such a structure is host-cell independent, ie, it is imposed by the protein primary structure of THP.⁴⁸ The percentage of distribution of high-mannose sequences in calf THP is very similar to that of human THP,⁴⁹ whereas pig THP has a much greater molar percentage of Man₅GlcNAc₂ (47% of total high-mannose sequences in pig THP versus 8% in human THP; Cavallone et al, manuscript submitted). The difference in the branching chain of high-mannose glycans between pig and human THP might affect the interaction with type 1 fimbriated *Escherichia coli*.⁵⁰ Recently, O-linked chains also have been described as present in THP.⁵¹

A question that has given rise to much controversy is whether the carbohydrate moiety of THP from pregnant and nonpregnant women differs.^{49,51-54} An accurate recent analysis⁵⁵ established that THP from pregnant women (also termed uromodulin) shows a small increase in Man₇GlcNAc₂ molar percentage (Man₇GlcNAc₂ represents 35% of total high-mannose glycans in pregnant women versus 30% in nonpregnant women). Conversely, neither in the course of pregnancy nor 1 month after gestation was a significant change observed in the negative charge distribution of complex-type glycans, indicating no change in the content of sialic acid and sulfate residues.

LOCALIZATION AND BIOSYNTHESIS OF THP

When RNAs isolated from approximately 150 different cell tissues were hybridized using a large probe for human THP RNA, only RNA from human adult kidney gave a positive signal, indicating that this glycoprotein is exclusively produced by kidney cells.¹³ Immunofluorescence and immunochemical analyses by light and elec-

tron microscopy indicated that THP resides in kidney cells of TAL and early distal convoluted tubules.⁵⁶⁻⁶¹ When the cellular location of rat THP messenger RNA (mRNA) was investigated by *in situ* hybridization using a radiolabeled human THP complementary RNA antisense probe (>600 bp), a positive reaction was found along the entire length of the TAL, but not in the macula densa or distal convoluted tubules.⁶² This last observation is in agreement with previous immunofluorescence analyses of hamster and human kidney performed in the laboratory of Robin D. Marshall,^{59,60} indicating that macula densa cells lack THP. Gokhale et al⁶³ used an RNA probe specific for rat THP and observed a strongly positive mRNA signal in the TAL region alone, even in conditions in which rat THP was detectable in papillary tubules. This result strongly supports the idea that rat THP synthesis is restricted to the TAL of Henle's loop. In a Western blot, we detected a relative distribution of human THP in kidney lysates of the outer medulla approximately 4-fold greater than in the cortex.⁴⁶

The first ultrastructural studies by Hoyer et al⁵⁷ and Seiler and Hoyer⁵⁸ failed to show in rats that THP is distributed preferentially at the luminal face of TAL cells. Subsequently, Bachmann et al,⁶¹ using protein-gold immunocytochemistry, observed a prevalent localization of rat THP in vesicles in close spatial relationship to the Golgi cisternae, fused with the apical face of TAL cells, and visualized slight positive gold labeling on the basolateral face. Conversely, various experimental approaches support the notion that THP is delivered to the luminal plasma membrane: (1) the GPI anchor and N-linked glycans both act as signals for vectorial transport of glycoproteins to the apical surface of epithelial cells,^{64,65} (2) prevalent exposure of THP at the luminal cell surface is consistent with both substantial excretion in luminal fluid and scarcity in blood,⁶⁶ and (3) recombinant THP expressed by Madin Darby canine kidney (MDCK) cells is delivered entirely to the apical face and released in the corresponding medium.⁶⁷

THP has been found in kidney distal tubules of one 14-week human fetus and in fetal rats,⁶⁸ whereas that found in human amniotic fluid very likely is derived from the fetus.⁶⁹

The biosynthesis of recombinant THP ex-

pressed by HeLa cells shows that THP accumulates intracellularly as a precursor of approximately 86 kd, which is converted into the mature glycoprotein with an apparent molecular weight of 97 kd (in reducing conditions). This mass shift depends on the processing of most high-mannose glycans to polyantennary species.⁴⁸ Two bands with a difference in migration similar to that of precursor and mature forms of recombinant THP have been visualized by Western blotting in lysates from human and rat kidney.^{30,70} When HeLa cells expressing THP are treated with mannosamine, an inhibitor of GPI anchor biosynthesis, THP accumulates intracellularly and is significantly less exposed at the cell surface, indicating that GPI anchor addition is required for delivery to the plasma membrane.³² Figure 3 shows the routing of GPI-anchored THP to the cell surface.

Both the first radioimmunoassay⁸ and the latest enzyme-linked immunosorbent assay (ELISA) method⁷¹ have shown that on consecutive days, there is a large variation in daily THP excretion under physiological conditions. Various parameters, such as urine volume and excretion of calcium, sodium, potassium, and creatinine, have been evaluated in relationship to daily THP urinary release, as have physical exercise, body weight, and diet, but controversial results have been obtained.⁷¹⁻⁷⁴ Moreover, the half-life for THP turnover in a single individual was reported to vary from a minimum of 3 to a maximum of 168 hours.⁷⁴ Recently, a significant increment in renal expression of THP in rats given a high-salt diet was observed.⁷⁵ Because THP urinary excretion is a multistep process that includes biosynthesis rate, complex posttranslational processing, and proteolytic release from the GPI-anchored counterpart, it is not surprising to find a broad scatter in the final step.

PROPERTIES OF THP

Tendency to Gelation/Aggregation

One of the most peculiar features of THP in solution is its tendency to gelation/aggregation when sodium chloride concentration is close to 100 mmol/L or calcium chloride is 1 mmol/L.^{76,77} Both conditions usually occur in normal urine, and a method based on this property has been set up to purify THP. When urine of healthy individuals is filtered through a diatomaceous

earth filter, THP is entrapped selectively in the filter. Because gelation is reversed at a low ionic concentration, THP is desorbed from the filter by deionized water and isolated to homogeneity by means of 2-step filtration and washing.⁷⁸ Recently, the salt precipitation method originally described by Tamm and Horsfall was compared with the diatomaceous earth filter method for isolating THP from urine of proteinuric patients and pregnant women, and better purification was obtained using the latter method.⁷⁹

Interaction of THP With Other Proteins

Many reports indicate that THP binds to proteins present in urine, particularly those occurring in pathological conditions, but the tendency to gelation/aggregation of THP at a sodium chloride concentration close to 0.1 mol/L might have been responsible for overestimation of the binding property of THP. A clear interaction between immunoglobulin G (IgG) and urinary THP was observed by Rhodes et al.^{80,81} They used a Scatchard plot to analyze the interaction of a monomeric form of THP, obtained by urea solubilization, to avoid the salt-dependent insolubilization of polymeric THP. In their first article,⁸⁰ these researchers reported a very high-affinity association between THP and IgG with a dissociation constant (K_D) ranging from 10^{-13} to 10^{-10} mol/L. In the second report, the same investigators corrected the K_D value, which actually ranged from 10^{-9} to 10^{-7} mol/L, but confirmed the biological significance of the interaction.⁸¹

Differences in the binding of IgG to THP have been observed in patients with glomerulonephritis or interstitial nephritis and related to changes in the carbohydrate moiety of THP, whereas in children with lymphoid cell malignancies, the alteration in THP glycoform does not affect interaction with IgG.^{82,83}

IgG light chains also bind THP, and when they are largely present in urine, as in patients with myeloma, this interaction is responsible for the cast nephropathy accompanying this disease⁸⁴ (see Cast Nephropathy). Both C1q and C1 bind THP with an affinity that decreases significantly when the electrolyte concentration of the THP solution shifts from 0.020 to 0.15 mol/L of sodium chloride.^{85,86}

Owing to the heterogeneity of the large THP glycoform, there are many observations about

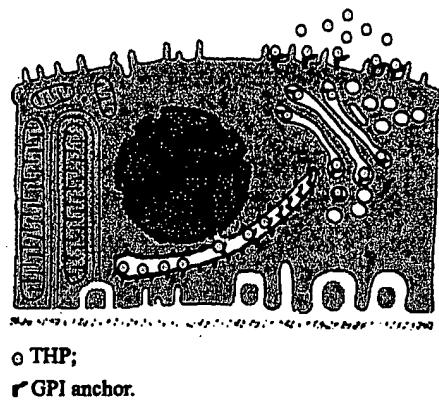


Fig 3. Schematic diagram of the biosynthesis and intracellular routing of GPI-anchored THP in a TAL cell. The addition of preformed GPI anchor to THP occurs in the ER: the membrane-bound form is transported to the Golgi complex, where glycans are fully processed, delivered to the luminal cell surface, and released in urine by a proteolytic cleavage (Fig 1). Note the large mitochondria and interdigitations in the basolateral membrane that are prominent in TAL cells.

its capacity for binding plant lectins, particularly those used as mitogens for lymphocyte blastogenesis, such as phytohemagglutinin (PHA), pokeweed mitogen, and concanavalin A.^{11,87} The strong inhibitory effect of urinary THP on PHA-induced lymphocyte blastogenesis is caused by the high-affinity interaction between PHA leucoagglutinin-subunits and tetra-antennary glycans, which are largely carried by THP.⁸⁸⁻⁹⁰ Very likely, the immunosuppressive activity assigned to THP from pregnant women (also termed uromodulin) depended on the competitive interaction of THP with PHA or pokeweed mitogen used in stimulating lymphocyte proliferation.^{10,14} Subsequent reports by the group of Muchmore^{52,91} showed that THP glycoform was entirely responsible for both in vitro inhibition of the antigen-specific lymphocyte proliferation and binding to recombinant interleukin-1 (IL-1) and recombinant tumor necrosis factor (TNF). According to Moonen and Williamson,⁹² soluble native cytokines do not bind to THP.

In our laboratory, we have shown that when high-mannose or complex-type THP glycopeptides are covalently bound to albumin, the neoglycoprotein inhibits the lymphocyte blastogenesis induced by 1-way mixed lymphocyte reaction. On this basis, we proposed that inhibition of lymphocyte proliferation is dependent on a multivalent interaction between THP glycans and

ligand(s) at the lymphocyte surface, and this interaction competes with the carbohydrate recognition system between effector and stimulator cells, which in the mixed lymphocyte reaction, mediates the blastogenesis.⁹³ Recently, it was reported that specific glycans of THP isolated from healthy adult males interact with IL-1 β , and glycans containing either N-acetylgalactosamine or sulfate residues in specific isomeric linkages are responsible for the inhibition of lymphocyte proliferation induced by tetanus toxoid.⁹⁴

Binding to and Activation of Leukocytes by THP

Several reports have shown that urinary THP binds to and activates leukocytes, including poly-

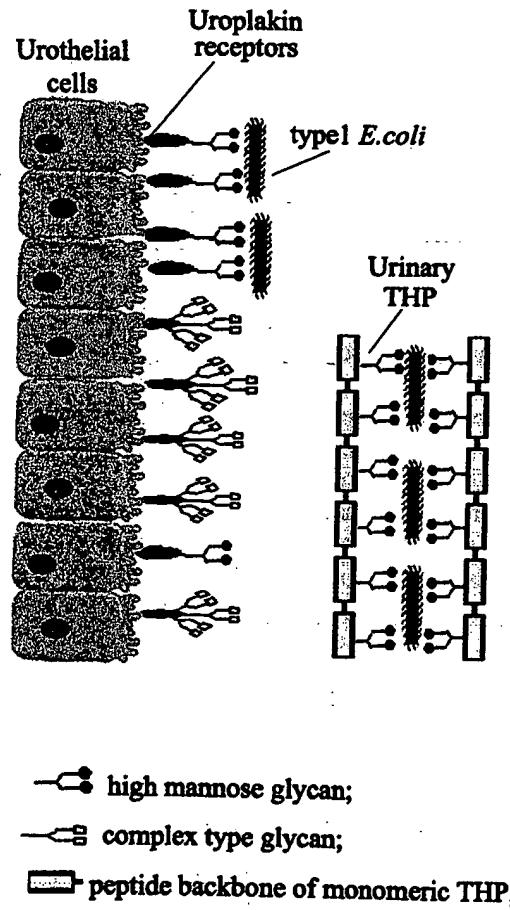


Fig 4. Binding of polymeric THP to type 1 *E. coli*, mediated by high-mannose glycans, competes with adhesion of pathogens to uroplakin receptors. Uroplakins are the major integral-membrane glycoproteins exposed at the luminal face of the bladder and urothelial tract. In the drawing of THP, polyantennary glycans have been omitted.

mophonuclear neutrophils (PMNs), lymphocytes, and monocytes. PMN activation is mediated by their interaction with THP (in monomeric form) through a single class of sialic acid-specific receptors exposed in great number at the surface of PMNs.^{95,96} Integrins also have been proposed to mediate the interaction between PMNs and THP through arginine-glycine-aspartic acid (RGD)-sequence mapping at position 142 to 144 of THP.⁹⁷ Recently, it was shown that recombinant THP produced by MDCK cells also binds to and activates PMNs.⁶⁷ According to Yu et al,⁹⁸ THP from pregnant women increases the phagocytic activity of PMNs and prostaglandin E₂ release, suggesting that a specific interaction occurs between THP and the membrane of PMNs. Similarly, blood mononuclear cells are activated by THP from pregnant women,⁹⁹ in this case, enhanced release of IL-1, IL-6, and TNF from monocytes very likely is responsible for the proliferation of both B and T lymphocytes. In a more recent study, Su et al¹⁰⁰ observed that THP from nonpregnant women also induces the secretion of TNF and expression of tissue factor by resting blood monocytes; however, both effects were several fold lower than those induced by THP from pregnant women (uromodulin).

Human monocytes isolated from peripheral blood and deprived of lymphocytes are able to phagocytize a particulate form of THP prepared from urine of healthy individuals and generate reactive oxygen metabolites, as well as release 95-kd gelatinase and other lysosomal enzymes.¹⁰¹ It is worth noting that the protection of mice against a lethal inoculum of *Listeria monocytogenes*, previously assigned to THP by Fontan et al,¹⁰² must be assigned to a minor protein (HGP92) also precipitated from normal urine by 0.58 mol/L of sodium chloride that shows an apparent molecular weight of 92 kd, very close to that of THP.¹⁰³

Binding of THP to Uropathogenic Strains of E coli

An interesting line of research concerns the role of THP in the kidney defense against urinary tract infections (UTIs), particularly those caused by *E coli*.¹⁰⁴⁻¹⁰⁷ UTIs occur in a large number of individuals (particularly women and children in developed countries), and the majority are caused by *E coli*.¹⁰⁸ Colonization is mediated by the

binding of lectin-like adhesins present on *E coli* fimbriae to carbohydrate structures carried by glycoproteins and glycolipids exposed at the cell surface. *E coli* fimbriae are classified according to their sugar specificity: type 1, type P, and type S recognize the high-mannose glycan, Galα1,4Galβ-sequence present in globoseries of glycolipids and the NeuAcα2,3Gal sequence of sialylated glycans, respectively.¹⁰⁹ Urinary THP carries both high-mannose and NeuAcα2, 3Gal sequences and thus may be considered a ligand for both type 1 and type S fimbriated *E coli*. There is evidence that type 1 *E coli* strains represent the predominant phenotypic variants of isolates from patients with UTIs,¹¹⁰ and uroplakin Ia and Ib (the most abundant integral membrane glycoproteins of the luminal surface in urothelial cells) behave as efficient cell receptors for type 1 fimbriated *E coli*.¹¹¹⁻¹¹⁴ Pak et al¹⁰⁷ recently showed that THP binds type 1 fimbriated *E coli* in vitro and efficiently competes with uroplakin Ia and Ib in binding to these pathogens. These results support the notion that in vivo, urinary THP represents a protective agent against UTI. Figure 4 shows the putative role of urinary THP in protection against UTIs.

CLINICAL RELEVANCE OF URINARY THP

Urinary THP is believed to be involved in the pathogenesis of various disorders of distal nephrons and the urinary tract, such as cast nephropathy, urolithiasis, and tubulointerstitial nephritis (TIN). Moreover, reduced urinary THP excretion is considered a reliable index of distal tubular cell damage. Finally, a very recent study showed that familial juvenile hyperuricemic nephropathy (FJHN) and autosomal dominant medullary cystic kidney disease 2 (MCKD2) arise from mutations of the THP gene.¹¹⁵

Cast Nephropathy

Microscopic observation of casts allows one to distinguish casts according to size and morphological characteristics. Although size reflects the shape of the tubules in which casts are formed, eg, they appear convoluted when formed in the convolute part of a distal nephron, a classification based on morphological characteristics consists of hyaline, granular, fatty, and leukocyte casts. THP is by far the predominant protein of hyaline casts, but also is present in the matrix of

casts in which plasma proteins or hemoglobin are abundant, as well as in granular casts containing debris from damaged kidney cells or blood cells.¹¹⁶

McQueen¹¹⁷ first proposed that THP was present in urinary casts. Subsequently, using a fluorescence antibody technique, he presented direct evidence of the widespread occurrence of THP in the matrix of urinary casts.¹¹⁸ Interestingly, in his first article, McQueen¹¹⁷ also reported that myeloma protein precipitates THP in aqueous solution. A few years later, Fletcher et al¹¹⁹ showed that THP isolated from casts of 4 patients with nephrotic syndrome and urine of healthy individuals showed a very similar amino-acid composition and carbohydrate content. More recently, Fairley et al¹²⁰ examined the protein composition of urinary casts from 60 patients with glomerulonephritis and 46 healthy individuals subjected to strenuous exercise. In addition to THP, plasma proteins, particularly IgA, IgG, or IgM, were found in the cast matrix of all patients, but THP was the only protein detected in hyaline casts from 41 healthy individuals, whereas in the remaining 5 individuals, fibrin, C3, and C1q, but not immunoglobulins, were present.

Recently, Fogazzi and Testanera¹²¹ reported that on the basis of histological examinations and chemical analyses of kidney and urinary casts, Carlo R. Rovida, who ran the Institute of Clinical Medicine at the University of Turin (Italy) from 1874 to 1877, first described a specific protein, termed cilindrina, produced by kidney tubular cells as the major constituent of hyaline casts.

The involvement of THP in hyaline cast formation is facilitated by its tendency to gelation/aggregation at a salt concentration close to isosmolarity.⁷⁶ It is worth noting that in the TAL, the tubular fluid in physiological conditions is at a low electrolyte concentration in that epithelial cells are not permeable to water, whereas chloride, sodium, and potassium are actively absorbed through Na,K-adenosine triphosphatase activity. Thus, in physiological tubular fluid in which THP is released from the GPI-anchored counterpart, the electrolyte concentration disfavors the gelation/aggregation of THP. There is evidence that removal of all N-linked glycans by N-glycanase treatment negatively affects the gel-forming tendency of THP.^{122,123}

The presence in urine of plasma proteins,

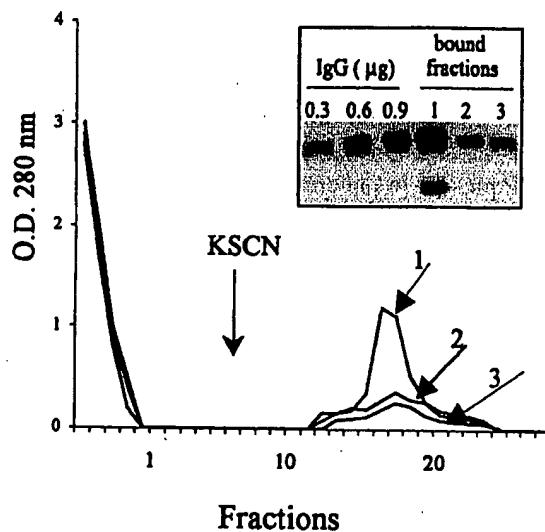


Fig 5. Representative profiles of affinity chromatography on a THP-CNBr-Sepharose column of urine from a patient with nephrotic syndrome (proteinuria, 3.7 g/L of protein). Arrows 1, 2, and 3 indicate the chromatography profile of urine dialyzed against 0.02, 0.08, and 0.15 mol/L of sodium chloride, respectively. (Inset) Western blot of pooled fractions containing bound proteins eluted by 3 mol/L of KSCN; lanes 1, 2, and 3 correspond to the 1, 2, and 3 chromatography. SDS-PAGE was performed in nonreducing conditions, and the blot was developed using anti-IgG antibodies.

particularly IgG and IgG light chains, is a crucial condition for cast formation. We observed that when IgG is excreted heavily by proteinuric patients, it binds to THP covalently linked to Sepharose in an amount that decreases significantly when sodium chloride concentration is increased from 0.02 to 0.15 mol/L. Figure 5 shows results of urine analysis from a patient with nephrotic syndrome (proteinuria, 3.7 g/L of protein). Similar results were obtained when proteinuric patients with other nephropathies were analyzed. These results support the notion that the electrolyte concentration of tubular fluid has a part in cast formation, particularly when IgG is present in urine.

Many studies^{84,124-127} reported the interaction between IgG light chains and THP. The THP sequence encompassing amino acids 225 to 233 appears to be crucial for binding to IgG light chains,¹²⁶ and the third complementary-determining region of both κ and λ light chains is required for binding to THP.¹²⁷ Leboulleux et al,¹²⁸ using an ELISA procedure, found that 5 of 12 samples of light chains from patients with cast nephropa-

thy did not react with THP and suggested that both a low affinity of certain light chains and differences in cast pathogenesis may explain these results. Consistent with the implication of THP in cast formation, GP2, a homologue of THP present in pancreas cells, is the major component of intraductal plugs of pancreatic juice.¹²⁹

Urolithiasis (Stone Disease)

Although the water-conservation function of the kidney produces supersaturation of various insoluble salts, such as calcium oxalate, calcium phosphate, and urate salts, only approximately 3% of the adult population of the Western hemisphere has nephrolithiasis.¹³⁰ The relatively low incidence of this disease depends essentially on 2 conditions: the presence in tubular fluid and urine of stone-formation inhibitors and the complex process required for the growth of microscopic crystal to a size producing stone disease. Stone formation usually takes place in various phases: the first consists of nucleation of supersaturated salts. Nucleation also may occur on heterogeneous surfaces present in tubular fluid or urine, eg, cell debris or urinary casts; subsequently, the microscopic nuclei undergo aggregation and grow into macroscopic stones.¹³⁰ Adherence of crystal aggregates to the tubular cell surface also is considered to have an important role in stone formation.¹³¹⁻¹³³

THP was found in human kidney stones in 1965 by Keutel,¹³⁴ and in 1973, Grant et al,¹³⁵ using a radioimmunoassay method, showed that THP content ranges from 0.002 to 5.07 mg/g of renal and bladder calculi, and there is no correlation between amount of THP and qualitative composition of inorganic components of stones.

The involvement of urinary THP in crystal nucleation has not been consistently proved,¹³⁶ whereas in an in vivo rat model, THP does not appear to mediate the initiation of crystal formation.¹³⁷ The inhibitory effect of THP in crystal aggregation and growth has been described in the case of both calcium oxalate¹³⁸⁻¹⁴⁰ and hydroxyapatite stones.¹⁴¹ There is consensus that inhibition of crystal growth in normal urine is caused mainly by urinary macromolecules, rather than low-molecular-weight components, and this property has been associated with the polyanionic structure of the major inhibitors.¹⁴² For instance, heparin and chondroitin sulfate are rich

in SO_4^{2-} groups,¹⁴³ whereas nephrocalcin, osteopontin (uropontin), and F1 activation peptide of prothrombin belong to the aspartic acid-rich protein superfamily.¹⁴⁴⁻¹⁴⁶ THP also is a polyanionic macromolecule (3.5 isoelectric point) because of the large extent of sialylation (18 residues/molecule according to Dustan et al⁹) and the presence of sulfate groups bound to *N*-linked glycans.³⁸ Recently, the presence in the luminal face of tubular cells of sialylated glycans carried by glycoproteins and glycolipids has been seen as crucial in the nucleation of calcium oxalate dihydrate crystals.¹⁴⁷ Conversely, adhesion of uric acid crystals to renal cells in culture (MDCK and BSC-1 line) was suggested to be mediated mainly by hydrogen bonds and hydrophobic interactions.¹⁴⁸ Because GPI-anchored THP is one of the main sialylated glycoproteins exposed at the luminal face of the first tract of distal nephrons, it should be involved in the calcium oxalate nephrolithiasis process even before it is released in urinary fluid. A recent study¹⁴⁹ reported decreased THP expression in the kidney of rats treated for 8 weeks with ethylene glycol to obtain calcium oxalate urolithiasis. However, reduced expression was detected only after 4 weeks of treatment, when crystal aggregates of calcium oxalate were visualized in the kidney of treated rats.

The difference in degree of sialylation between urinary THP from healthy individuals and recurrent calcium oxalate renal stone formers has been investigated extensively. Some studies¹⁵⁰⁻¹⁵³ reported an increase in isoelectric point of THP from recurrent calcium oxalate stone formers, probably dependent on a reduced sialic acid content. However, this observation was not confirmed subsequently.^{63,154} Despite this, partial removal of sialic acid from THP by neuraminidase treatment results in loss of in vitro inhibition of urinary crystal aggregation,^{155,156} consistent with the notion that polyanionic structure is crucial for inhibition of calcium oxalate crystal aggregation.

The involvement of urinary macromolecules in the modulation of crystal aggregation, as well as their participation in stone formation, was analyzed in some studies by using diluted urine or urine processed by centrifugation or filtration. The last 2 procedures, particularly filtration, significantly reduced THP content in processed

urine.¹⁵⁷ It is evident that this behavior may have been responsible for discrepancies in results concerning the role of THP as a modulator of urolithiasis, particularly when this role was compared with that of other urinary modulators that do not significantly change concentration on urine processing.

TIN

Bacterial infections, vesicoureteral reflux, exposure to heavy metal, and antibiotic or analgesic intake are the major etiologic agents of TIN, which is characterized in the first phase by infiltration of inflammatory cells (mononuclear cells, lymphocytes, plasma cells, and PMNs) into kidney interstitium and subsequently by interstitial fibrosis and tubular atrophy.¹⁵⁸ Clinical studies indicated that renal diseases, namely, chronic interstitial nephritis, medullary cystic disease, reflux nephropathy, and rejecting renal allografts, very often are accompanied by abnormal THP deposits in the tubular interstitium, and there is an immunologic response to THP.¹⁵⁹⁻¹⁶³ THP is a powerful autoantigen, and immune deposits containing THP and anti-THP antibodies have been localized in the intercellular space of TAL in rats, mice, and rabbits challenged with homologous THP.^{58,164-167} In all likelihood, healthy mammals, including humans, do not produce anti-THP antibodies because the exclusive localization of THP at the luminal face of tubular cells segregates the protein from the immune system. Segregation may be abolished by various alterations to cells expressing THP, such as loss of their apical/basolateral polarity and the consequent release of THP from the wrong side, as well as conditions altering cell integrity. Nonvectorial delivery of THP to the luminal surface also may occur in conditions that alter the routing of THP along the exocytic pathway. Malagolini et al³² observed that under treatment with monensin, an ionophore that interferes with the function of the Golgi apparatus, a THP that is not fully glycosylated and not exposed at the cell surface is released from cells expressing THP. Because the GPI anchor is a sorting signal to the cell apical face,⁶⁴ if the GPI anchor is lost along the exocytic pathway, THP also might be released from the basolateral face and enter the peritubular capillaries; tubular basement mem-

branes are highly permeable to macromolecules.¹⁶⁸

Several studies have shown that interstitial deposits of THP and THP-immune complexes frequently are surrounded by neutrophils, mononuclear cells, and plasma cells.¹⁵⁹⁻¹⁶⁴ Although binding of THP to neutrophils may be responsible for the acute inflammatory response,⁹⁵⁻⁹⁷ binding to mononuclear cells may extend the reaction to the chronic phase characterized by fibrosis.^{101,165} Although the proinflammatory activity of THP has been put forward as involved in TIN pathogenesis,^{96,101} both clinical and experimental observations indicate that abnormal THP deposits in the kidney do not have a crucial role in inflammatory processes. For instance, on the basis of 124 renal biopsies of patients with various nephropathies, Chambers et al¹⁶⁹ concluded there is no correlation between THP accumulation in the interstitium and tubulointerstitial damage, although in the majority of patients examined, THP antibodies were present. The absent or low inflammatory response to extratubular THP deposits (particularly PMN infiltration) also was reported in renal allografts and patients with urinary extravasation.¹⁷⁰⁻¹⁷² Again, in experimental models of unilateral ureteral obstruction in rats or mice, as used by Dziukas et al¹⁷³ and Fasth et al,¹⁷⁴ THP accumulation in the interstitium is not accompanied by inflammatory response. These investigators suggested that in urine extravasation, toxin components other than THP may be responsible for the inflammation and TIN pathogenesis.

When transfected HeLa cells stably expressing cell surface-anchored THP were incubated with human neutrophils, no binding was found, but opsonization of cells with anti-THP antibodies resulted in (1) dramatic adhesion and myeloperoxidase activation of PMNs, (2) a significant increase in THP release, and (3) exit of the partially processed THP from the cells. These results support the notion that after the autoimmune response, cell surface-anchored THP also may contribute to the pathogenesis of TIN.¹⁷⁵

Involvement of THP in tubular cell damage also has been related to its ability to bind to C1q, the component of the complement pathway responsible for initiating the complement cascade.^{85,86} There is evidence that in some renal diseases (eg, nephrotic syndrome), complement

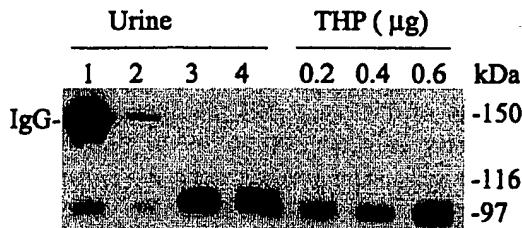


Fig 6. Western blot of urine samples from patients with lupus nephritis (lane 1) and MCKD (lane 2) and 2 healthy individuals (lanes 3 and 4). An identical percentage of daily urinary volume (0.5%) was applied in each lane. Urinary THP used as a marker was isolated from pooled urine samples of healthy individuals. SDS-PAGE was performed in nonreducing conditions, and the blot was developed using both anti-THP antibodies and anti-IgG antibodies. As expected, the latter antibody visualized a large presence and a trace of IgG in urine from patients with lupus nephritis and MCKD, respectively, and absence of IgG in healthy individuals.

factors are excreted copiously and activated in urine.¹⁷⁶ Thus, either complement per se or the THP-C1q complex may accumulate in the interstitium and have an important role in TIN pathogenesis.

THP Excretion in Nephropathies

Abnormalities in daily urinary excretion of THP have been detected in several pathological states of the kidney and urinary tract, and decreased excretion is considered a reliable index of damage to the TAL cells that synthesize THP.^{73,177,178} Our laboratory detected a marked reduction in daily THP excretion in a patient with medullary polycystic kidney disease (Fig 6). In active lupus nephritis, decreased THP excretion was related to impairment of the distal nephron function that accompanies the active phase of the disease¹⁷⁹ (Fig 6). Again, marked reduction in urine THP from infants with hyperprostaglandin E syndrome was related to a congenital defect of the distal nephron.¹⁸⁰ THP excretion also is reduced in urine of stone formers, very likely because stones per se damage the tubular epithelium of the TAL.¹⁸¹⁻¹⁸³ Interestingly, a study¹⁸⁴ of THP excretion after uninephrectomy in kidney donors has shown a persistent increase (~40%) in THP excretion from the kidney that remains in the donor and good correlation between THP excretion rate and glomerular filtration rate.

Urinary THP has been studied in diabetes mellitus, particularly the insulin-dependent

type.¹⁸⁵⁻¹⁹³ A reduction in THP excretion was observed consistently when diabetic nephropathy is characterized by a reduction in renal mass, which usually occurs at a late stage of disease.¹⁸⁵⁻¹⁹⁰ Conversely, other investigators observed increased excretion in the early stage of the disease, as well as during acute euglycemia and water load.¹⁹¹⁻¹⁹³ In experimental diabetes induced in rats by streptozotocin, THP excretion was increased markedly, an effect supposed to be associated with distal tubular dysfunction.^{194,195} According to Agardh et al,¹⁹⁵ greater excretion is attenuated by compounds that inhibit both the production of advanced glycation end products and oxidative stress, 2 processes involved in the development of diabetic nephropathy.

Mutations to the THP Gene in FJHN and MCKD2

The autosomal dominant MCKD2 and FJHN, very likely 2 facets of the same disease, are characterized by hyperuricemia, medullary cysts, interstitial nephritis, and progressive renal failure.¹⁹⁶ By genome-wide linkage mapping in Italian, Czech, and Belgian families, new loci for MCKD2 and FJHN were identified in the regions of chromosome 16p11.2 and 16p12,¹⁹⁶⁻¹⁹⁸ close to the localization of the human THP gene.²¹ Mutations of the THP gene were identified very recently by Hart et al¹¹⁵ in several individuals from 3 unrelated families with FJHN and 1 family with MCKD2. The missense mutations identified consist of the deletion or addition of 1 cysteine residue and 1 frame deletion of 9 amino acids. These results provide the first demonstration that FJHN and MCKD2 arise from allelic mutations of the THP gene and suggest that an alteration to the tertiary structure of THP, particularly the misfolding caused by the uncoupling of at least 1 Cys-Cys bond, is responsible for the disease(s).

We observed³² that when HeLa cells expressing THP are treated with an exogenous reducing agent, such as 2-mercaptoethanol, routing along the exocytic pathway of partially reduced THP is delayed considerably, indicating that completion of intrachain disulfide bonds is required for a regular exit from the ER and delivery of THP to the cell surface. On this basis, one may postulate that misfolded THP produced in MCKD2 and FJHN undergoes a similar fate, and accumula-

tion in the ER may result in altered biosynthesis, eg, impairment of the GPI addition and glycosylation processing, 2 events crucial for the delivery of THP to the luminal surface of the tubular epithelium. These effects may explain the TIN associated with the disease(s).

THP IN BIOTECHNOLOGY

Observations that the THP gene is transcribed exclusively by TAL cells and THP is largely excreted in urine recently have been used for the transgenic production of human therapeutic proteins.^{199,200} This approach is based on recently developed technologies that allow one to generate transgenic mice by using a mouse gene promoter to direct the expression of proteins selectively released in urine.²⁰¹ In comparison to the expression of human therapeutic proteins in milk of transgenic livestock, production by kidney cells and the consequent release in urine appears to offer a much more cost-effective advantage in that urine contains few proteins or lipids; a condition that facilitates isolation of the recombinant protein to homogeneity. In this way, recombinant human α_1 -antitrypsin was isolated from transgenic mice. It shows a broad similarity in its activity to that produced by human hepatocytes.¹⁹⁹ When the same approach was used to produce human recombinant erythropoietin, its expression and excretion in transgenic mice was accompanied by disease symptoms similar to polycythemia in humans, confirming that the THP promoter is effective in directing the production of proteins capable of exerting an *in vivo* effect.²⁰⁰

Recently, research in the field of genomic manipulation acquired a new strategy, referred to as the Cre/loxP site-directed recombinant system.²⁰² This strategy provides an advantageous tool for spatially and temporally modulated somatic mutations. When stable transgenic mice expressing Cre recombinase, under a tissue-specific promoter, are mated with mice harboring the loxP DNA sequence, a new mouse line may be obtained in which the gene disruption mediated by Cre recombinase occurs only in cells in which the promoter is active.²⁰² Striklett et al²⁰³ used this strategy and, using a THP promoter, found that Cre-recombinase is specifically expressed and active in cells of TAL. As these investigators emphasized, this approach intro-

duces a new tool in specifically exploring the function of the TAL, which represents one of several regions of the kidney that variously contribute to renal physiological states.

CONCLUSION

Although the bulk of evidence shows the involvement of THP in pathological states of the kidney and urinary tract, there are no clear indications about its physiological role, although the structural, genetic, and cytological characterization largely has been clarified in the last 20 years. In the late 1980s, great enthusiasm accompanied the observation that urinary THP/uromodulin could have an immunoregulatory function in that it inhibits *in vitro* lymphocyte blastogenesis and interacts with cytokines.²⁰⁴ Some considerations may be proposed to explain why this line of research has not confirmed initial expectations and hopes for the therapeutic use of THP. First, results obtained *in vitro* have not been carefully evaluated with respect to their *in vivo* relevance; in physiological conditions, THP is expressed exclusively by TAL cells, delivered to the luminal surface, and excreted in urinary fluid. Thus, it is improbable that a protein present in a cell compartment and fluid, where very few cells mediating the immune response occur, may show an immunoregulatory function.

The second consideration concerns the demonstration that the glycoform of THP is entirely responsible for inhibiting lymphocyte proliferation and cytokine binding.^{52,91} Because completion of the carbohydrate moiety of a glycoprotein is dictated specifically by enzymes (glycosidases and glycosyltransferases) from cells responsible for expressing it,²⁰⁵ large-scale production of a functionally active THP cannot be made by the usual cost-effective biotechnological techniques. Last, but not least, the property by which THP behaves as an efficient autoantibody prevents it from being used *in vivo*.

Considering that release of THP in the urinary fluid from the cell-surface GPI-anchored counterpart is relatively constant and abundant in physiological conditions and all mammals excrete THP, one may assume that its presence in urine is an advantage very likely imposed by selective pressure. Thus, the question is, what condition(s) served as a selective agent for this process? One possible answer is that urinary THP affords effi-

cient protection toward the most frequent infections of the urinary tract by binding uropathogenic strains of *E. coli*.¹⁰⁷ The luminal cell surface of the respiratory and intestinal epithelium is covered by mucus, an effective agent in preventing adhesion of pathogens to glycoproteins and glycolipids exposed at the luminal plasma membrane. This protection is absent from the urinary tract in that no mucus (or only a very small amount) covers the luminal cells, particularly in the bladder, in which urine constantly resides. Because infections are caused mainly by *E. coli* strains that enter the urinary tract by an ascending route from intestinal flora,²⁰⁶ a real advantage seems to be offered by the presence of THP as a soluble ligand because, following the route and fate of urine, it may interact with the pathogens any time after their entry into the urinary tract and eventually be eliminated from the body. The occurrence of THP in urine as a large homopolymer also is a very advantageous property in that it behaves as a multivalent ligand, ie, it binds pathogens with a high affinity. In this context, addition of the GPI anchor also may be an advantage from an evolutionary point of view because the GPI anchor is a selective signal for delivering glycoproteins to the luminal cell surface of tubular epithelium and for the restricted release of THP into the urinary fluid.

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